The CH50 assay consists of analysis of complement-dependent lysis of sheep red blood cells (RBCs) which have been opsonized with anti-sheep RBC serum and is dependent on both Mg^{++} and Ca^{++}. The CH50 is the concentration of human serum necessary to cause the lysis of 50% of the opsonized sheep RBC within 1 hour at 37°C. The primary screen for C2 inhibitors includes use of a constant serum concentration at the CH50 level, and the assay is conducted in the presence and absence of 10 μM of test compounds. Compounds that inhibit this primary assay are titrated and retested for specificity in a secondary hemolytic assay in which each individual purified complement protein is added sequentially in the presence or absence of the test compound to determine which component is being inhibited.

The AH50 assay consists of analysis of the direct complement-dependent lysis of rabbit red blood cells and is dependent on Mg^{++} but not Ca^{++}, and therefore is performed in the presence of EGTA. Similar to the CH50, the AH50 is the concentration of human serum necessary to cause the lysis of 50% of the rabbit RBC within 1 hour at 37°C. The primary screen for Factor B inhibitors includes use of a serum concentration at the AH50 level, and the assay is conducted in the presence and absence of 10 μM of test compounds. Compounds that inhibit this primary assay are titrated and retested for specificity in a secondary hemolytic assay in which each individual purified complement protein is added sequentially in the presence or absence of the compound to determine which component is being inhibited.

Sheep whole blood in Alsevers solution and anti-sheep hemolysin were obtained from Colorado Serum Co. (Denver, CO). Erythrocyte-antibody complexes (EA) were produced using an optimal concentration of anti-sheep hemolysin, determined by titration to be a 1:800 dilution. Normal human serum (NHS) was generated by collecting fresh serum from 10 random healthy human donors, pooling it, aliquotting the pooled serum, flash freezing it in liquid nitrogen, and storing it at −70°C. A fresh aliquot was thawed immediately prior to each use.

A standard assay was established in Costar 96-well round-bottom or V-bottom microtiter plates. All samples were analyzed in duplicate and averaged. First, the NHS was titrated to determine the midpoint of its linear activity in lysing the EA

(the CH50 dilution). Serial two-fold dilutions of freshly thawed NHS in gelatin-veronal buffer with Mg^{++} and Ca^{++} (GVB^{--} containing 0.142 M NaCl, 4.9 mM sodium 5, 5'-diethylbarbituric acid and 1.0 g/l gelatin, the pH adjusted to 7.35 with HCl, followed by addition of $CaCl_2$ and $MgCl_2$ to final concentrations of 60 μM and 400 μM, respectively), or dH_20 (used to determine total lysis) were placed in duplicate wells (80 μl/well) and warmed to 37°C for 5 minutes. EA which had been washed twice with GVB^{++} and resuspended at 2 x 10^8 complexes/ml were added (80 μl/well) and the plate was incubated at 37°C for 60 minutes. Eighty μl/well of 0.15 M NaCl was added and the plate was centrifuged at 2500 rpm for 3 minutes. One hundred μl/well of supernatant was transferred from the assay plate to an Immulon4 96-well flat-bottom ELISA plate and the absorbance at 420nm was determined. Background readings of absorbance in the wells containing no NHS were subtracted from the reading for all wells containing NHS and the resulting specific absorbance was expressed as a percentage of that obtained from wells containing dH_20 (% Total Lysis).

The dilution of NHS necessary to give 50% Total Lysis in 60 minutes at 37°C (the CH50) was determined to be 1:150. This dilution constituted the midpoint of the linear range of the NHS lytic activity and was used to screen the library of test compounds for inhibitors of the complement pathway. The test compounds were first diluted in GVB^{++}/5% DMSO to 40 μM and aliquotted at 40 μl/well in duplicate into Costar 96-well round-bottom or V-bottom plates. Control wells containing GVB^{++} (background), dH_20 (total lysis), DMSO alone, anti-C2 polyclonal antisera (40 μg/ml; Calbiochem), normal goat IgG (40 μg/ml; Sigma), and EGTA (4 mM) were also included. Plates were incubated at 37°C for five minutes. Forty μl/well of NHS diluted to 1:75 in GVB^{++} was added (this created a 1:150 final dilution with compound), except in background or total lysis wells which received GVB^{++} or dH_20, respectively. Plates were incubated at 37°C for 10 minutes. EA were washed twice, resuspended at 2 x 10^8/ml in GVB^{++}, and added to each plate at 80 μl/well. The plates were incubated at 37°C for 60-70 minutes, after which 80 μl/well of 0.15 M NaCl was added and the plates were centrifuged at 2500 rpm for 3 minutes. One hundred μl of supernatant from each well was transferred from the assay plates to

separate wells on Immulon4 96-well flat-bottom ELISA plates and the absorbance at 420nm was analyzed. Background readings of absorbance in the wells containing no NHS were subtracted from the absorbance for each well and the resulting specific absorbance was expressed as a percentage of that obtained from wells containing DMSO alone (% DMSO lysis). All compounds which inhibited DMSO lysis by greater than 35% were re-tested and titrated in the same assay. Thirty nine compounds were identified with IC50 values of less than or equal to 20 μM. The two most potent compounds had IC50 values of less than 5 μM, and were shown to be selective for complement inhibition since they did not significantly inhibit (i) LFA-1 mediated adhesion to ICAM-1, (ii) Mac-1 mediated adhesion to ICAM-1, (iii) $a_2\beta_1$ mediated adhesion to collagen, (iv) $a_4\beta_7$ mediated adhesion to MAdCAM-1, or (v) vWf binding to gp1b in standard cell-based adhesion assays at concentrations greater than or equal to 20 μM.

Approximately 30% of the activity of serum in the classical complement pathway (CCP) screen is due to amplification by the alternative complement pathway (ACP) Factor B containing C3 and C5 convertases. Therefore, this assay has the potential to isolate inhibitors of either the classical complement pathway convertases, the lectin complement pathway (LCP) (in which C3 is an intermediate component as well), and the alternative complement pathway. It is also possible that given the high degree of primary structural homology between C2 and Factor B, compounds may be isolated which inhibit both convertases in all three pathways.

Given the nature of the original screen, inhibition could have occurred at any stage of the complement pathway. In order to determine at which stage of complement activation the test compounds inhibited activity, purified complement proteins were obtained (Advanced Research Technologies, San Diego, CA) and complement activation was reconstituted in a stepwise manner. At each step, the lead compound or DMSO alone was added and the terminal hemolytic activity was measured as above. Initially, the lead compound was tested for its ability to inhibit at any of four different stages of complement activation: 1) C1 binding to aggregated antibody on the surface of the EA; 2) C4 binding to and cleavage by C1; 3) C2

binding to C4b, activation of C2 by C1-mediated cleavage and C4bC2a-mediated cleavage of C3 (i.e., formation and activity of the C3 convertase); and 4) formation and activity of the C5 convertase and subsequent deposition of complement proteins C6 through C9, which form the membrane attack complex (MAC) resulting in cell lysis.

In the assay, 1 x 10^7 EA/well were analyzed in duplicate wells of Costar 96-well round-bottom plates. For testing stage 1 (as indicated above), cells were resuspended in GVB^{++} containing 7.5 µg/ml C1 protein and incubated for 15 minutes at 30°C. For testing stage 2, cells were resuspended in GVB^{++} containing 7.5 µg/ml C4 protein and incubated for 15 minutes at 30°C. For testing stage 3, cells were resuspended in GVB^{++} containing 0.4 µg/ml C2 protein and 25 µg/ml C3 protein and incubated for 30 minutes at 30°C. For testing stage 4, cells were resuspended in GVB^{++} containing 4 mM EGTA and a 1:50 dilution of NHS and incubated for 60 minutes at 37°C. For each stage, a titration of the lead compound was carried out wherein the dilutions of the compound with DMSO, goat anti-C2 pIgG, and goat normal pIgG were tested for inhibition. Each pair of wells received inhibitors at only one stage. After each stage's incubation period, plates were centrifuged at 2400 RPM for 3 minutes, and cell pellets were washed twice with 100 µl/well GVB^{++} to remove inhibitors and unbound protein. EGTA was used in stage 4 to block new addition of C1 from the serum and therefore make the final stage dependent on previous deposition of C3b. In this component assay, anti-C2 pIgG but not normal pIgG, blocked complement activation at stage 3 as expected.

The lead compound inhibited stage 4 in a dose-dependent manner but not stages 1, 2, or 3. These results indicated that the compound did not inhibit formation or activity of the CCP/LCP C3 convertase but inhibited either the C5 convertase or subsequent formation of MAC, the terminal component of the complement system.

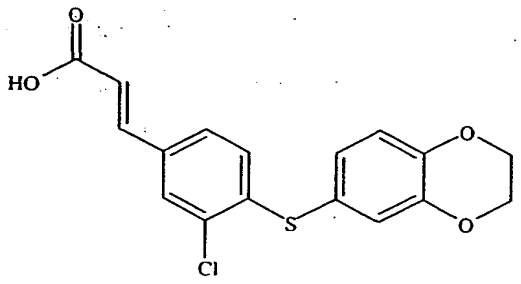
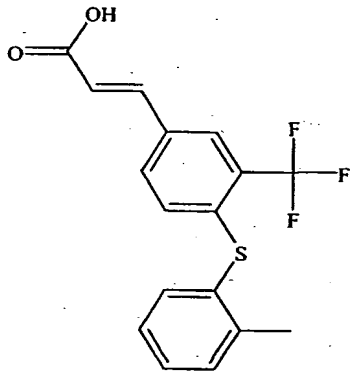
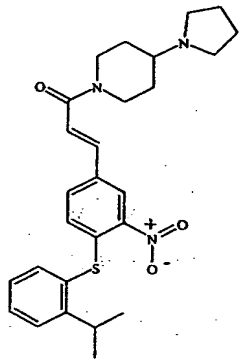
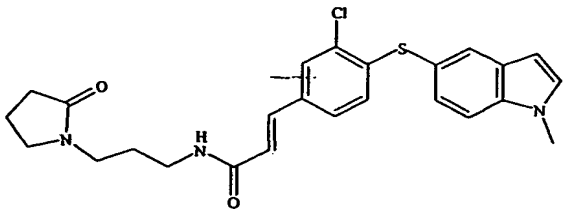
In order to determine whether the lead compound inhibited the activity of the C5 convertase or subsequent formation of the MAC, a simplified component assay was carried out. C2-depleted NHS was obtained (Advanced Research Technologies, San Diego, CA). EA were washed twice with GVB^{++} and resuspended

at 2 x 10^9 cells/ml in GVB^{++}. An equal volume of GVB^{++} containing a 1:50 dilution of C2-depleted NHS was added and the cells were incubated at 30°C for 7.5 minutes to allow deposition and activation of C1, and subsequent cleavage of C4. The cell suspension was diluted 20-fold with GVB^{++} to stop the reaction, and centrifuged 2400 RPM for three minutes. The cell pellet was washed three times with GVB^{--} and resuspended at 2 x 10^8 cells/ml in GVB^{++}. Fifty µl/well of the treated EA was added to duplicate wells of a Costar 96-well round-bottom plate, along with 50 µl/well of GVB^{++} containing 1 µg/ml C2, 50 µg/ml C3, and 1 µg/ml C5, with or without anti-C2 (80 µg/ml) normal goat IgG (80 µg/ml). Lead compound (80 µM) or DMSO was added and the plate was incubated at 30°C for 20 minutes. Two hundred µl/well of GVB^{++} was added, the plate was centrifuged 2400 RPM, 3 minutes, the supernatants were aspirated, and the pellets washed once with 200 µl/well GVB^{++}. The cell pellets were resuspended in 100 µl/well GVB and 100 µl/well GVB containing 40 mM EDTA and 1:50 NHS was added, after which the plate was incubated at 37°C for 60 minutes. The plate was centrifuged again, and 100 µl/well was transferred to an Immulon4 96-well flat-bottom plate and absorbance determined at 420 nm.

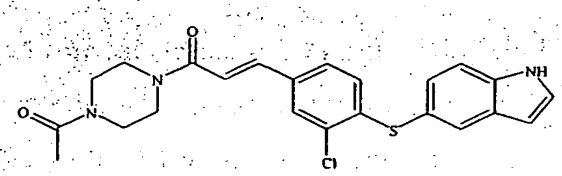
Both the anti-C2 pIgG and the lead compound specifically inhibited hemolysis, indicating that the compounds inhibit the CCP/LCP C5 convertase activity directly. These results were consistent with a potential mechanism of complement inhibition wherein the test compound bound C2 or Factor B and inhibited a conformational change necessary for the serine protease domain to gain access to the C5 substrate. Crystal structure data of the Factor B serine protease domain and modeling of its interaction with the A domain is consistent with this hypothesis [Hua Jing, *et al.*, EMBO J. 19:164-173 (2000)].

The top 5 inhibitors of complement proteins C2 and Factor B are shown in Table 4.

TABLE 4

AO	
AP	
AQ	
AR	

AS



Example 5
Isolation of cDNAs for Alpha E, E-cadherin, and MAdCAM-1

In order to assess whether it is possible to modulate binding activity of other α/β proteins, DNA encoding alpha E, E-cadherin and MAdCAM-1 were prepared as follows.

A. Alpha E

1. Isolation of human alpha-E cDNA

DNA encoding human alpha-E was isolated from a normal human intestinal cDNA library (Clontech Laboratories, Inc., Palo Alto, CA) using an alpha E I domain cDNA as a probe. The alpha E I domain probe was cloned by PCR amplification using a human colon cDNA library as template and primers encompassing the 5' and 3' ends of the alpha E I domain. In order to facilitate cloning, *Bam*HI and *Xho*I restriction sites (underlined in the sequence) were designed into the 5' (SEQ ID NO: 7) and 3' (SEQ ID NO: 8) primers.

ATT GGA TCC GCT GGC ACC GAG ATT GCC ATC — SEQ ID NO: 7
AAT TTC TC GAG GTC TCC AAC CGT GCC TTC C — SEQ ID NO: 8

A 607 bp I domain fragment was amplified, digested with *Bam*HI and *Xho*I, and inserted into the plasmid pBluescript® SK (Stratagene, La Jolla, CA). The plasmid was transformed into bacteria, plasmid DNA was prepared according published procedures, and the *Bam*HI/*Xho*I insert was purified. The fragment encoding the alpha E I domain was radiolabeled with ^{32}P-dCTP and ^{32}P-dTTP using a random primed DNA labeling kit (Roche Diagnostics Corp., Indianapolis, IN) for use as a hybridization probe.

DNA encoding full-length alpha E was identified as follows. A human intestinal cDNA library in phage lambda GT11 (CLONTECH Laboratories, Inc., Palo Alto, CA) was plated and hybridized with the I domain probe using standard procedures. From two rounds of screening, six phage clones were isolated. The cDNA inserts were isolated from the phage by *Eco*RI digestion, subcloned into

pBluescript® SK (Stratagene, La Jolla, CA), and sequenced. A complete 3.4 kb sequence was reconstituted from three different clones: clone A (3) encompassing the 5' end, clone B (22) that included sequences from in the middle of the cDNA, and clone C (22) encompassing the 3' end of alpha E cDNA. Sequence analysis indicated that clone A (3) contained an insertion of two cytidines and another insertion of a guanine at positions 357 and 464, respectively, when compared to the published nucleotide sequence. These insertions resulted in a 75 base frameshift in the open reading frame which resulted in the addition of 25 additional amino acid residues, shown below, not found in the previously reported sequence.

PKGRHRGVTVVRSHHGVLICIQVLVRR SEQ ID NO: 9

The sequences downstream from this 25 amino acid insertion were identical to the published alpha E sequence for the rest of the molecule.

In order to subclone the alpha E cDNA into pcDNA3® (Invitrogen Corp., Carlsbad, CA), a *Hin*dIII site was generated at the 5' end by PCR amplification using the 5' primer Eo26-H3 (SEQ ID NO: 10) and the 3' primer Eo-24 (SEQ ID NO: 11) primers shown below.

GAG GGG AAG CTT AGT GGG CC SEQ ID NO: 10
GAA GTT GGC CTG AGC CTG G SEQ ID NO: 11

The PCR product was digested with *Hin*d III and *Nsi*I, and ligated into the corresponding sites of the vector.

The expression vectors pMHneo [Hahn *et al.*, Gene 127:267-268 (1993)] and pcDNA3®/aE were transformed into the bacterial strain NEB316, a dam⁻ strain which does not methylate *Xba*I restriction sites, and plasmid DNA isolated according to standard procedures. Both pMHneo and pcDNA3®/aE were digested with *Hin*dIII and *Xba*I and the 3.4 kb alpha E cDNA fragment from pcDNA3®/aE was separated using agarose gel electrophoresis. The fragment was excised from the gel, purified, and ligated into *Hin*dIII/*Xba*I-digested vector pMHneo. An aliquot of ligation mixture was used to transform XL-1 Blue bacteria (Stratagene, La Jolla, CA)

according to the manufacturer's protocol, and bacterial colonies containing pMHneo were selected by growth on LBM agar plates containing ampicillin. Bacterial colonies were grown overnight in LBM media containing 100 ug/ml ampicillin and plasmid DNA was isolated using the Wizard Plus Miniprep Kit (Promega Corp., Madison, WI). The plasmid DNA was characterized by diagnostic restriction digestion and a plasmid containing the alpha E cDNA, referred to as pMHneo/aE, was used to stably transfect a JY cell line as described below.

B. E-cadherin

1. Isolation of E-cadherin cDNA

The cDNA for human E-cadherin was isolated by PCR amplification of a Marathon-Ready™ human colon cDNA library (CLONTECH Laboratories, Inc. Palo Alto, California) using E-cad 5'#1 (SEQ ID NO: 12) and E-cad 3'#1 (SEQ ID NO: 13) primers, which are set forth below.

5'-CTGCCTCGCTCGGGCTCCCCGGCCA-3'	SEQ ID NO: 12
5'-CTGCACATGGTCTGGGCCGCCTCTCTC-3'	SEQ ID NO: 13

Polymerase chain reactions were performed in a Perkin Elmer Cetus (PE Applied Biosystems, Foster City, CA) DNA thermal cycler in a reaction mixture containing 5 µl of the library cDNA, 10 µl of 5X PCR buffer from an Advantage™-GC cDNA PCR Kit (CLONTECH Laboratories, Inc. Palo Alto, California), 1 µl of 50X dNTP mix, 1 µl of 10 µM primer E-cad5'#1, 1 µl of 10 µM primer E-cad 3'#1, 1 µl of Advantage™ KlenTaq polymerase mix, and 31 µl of H_2O. Amplification conditions included an initial incubation for 1 min at 94°C, followed by 5 cycles at 94°C for 30 sec and 72°C for 4 min; 5 cycles at 94°C for 30 sec and 70°C for 4 min; 25 cycles at 94°C for 30 sec and 68°C for 4 min; and a final 5 min incubation at 72°C. An aliquot of the reaction was separated using agarose gel electrophoresis to determine the approximate size of the PCR product and a single band of ~2.7 kb was detected as anticipated. The 2.7 kb PCR product was ligated into the plasmid pCR®2.1 using a TA Cloning® Kit (Invitrogen Corp., Carlsbad, California) according to the manufacturer's protocols. *E. coli* strain INVaF' (Invitrogen Corp., Carlsbad, CA) was

transformed with an aliquot of the ligation reaction as recommended by the manufacturer and single bacterial colonies were isolated and grown overnight in LBM media containing 100 µg/ml ampicillin. Plasmid DNA was isolated from these cultures using the Wizard Plus Miniprep Kit (Promega Corp., Madison, WI).

2. Generation of DNA encoding a E-cadherin/Ig fusion protein

The extracellular region of E-cadherin is made up of five tandem repeats (domains) of approximately 110 amino acids each. In order to express an E-cadherin-human/human IgG1 fusion protein, a DNA fragment containing domains 1 through 5 of E-cadherin was generated by PCR amplification of the E-cadherin cDNA (pCR®2.1/E-cadherin #3 described above) with primers Ecad5'Kozak (SEQ ID NO: 14) and Ecad3'(Xho) (SEQ ID NO: 15). The 5' primer Ecad5'Kozak was used to add a 5' *Hin*dIII site to facilitate subsequent subcloning of the 5-domain fragment into the expression vector pDEF2 (see U.S. Patent 5,888,809) and reconstitute a Kozak sequence upstream of the translation initiation codon which was lacking from initial E-cadherin cDNA clone. The 3' primer Ecad3'(Xho) generated a new 3' end of the fragment containing domains 1 through 5 of E-cadherin, and added a *Xho*I restriction site to the 3' terminus of the fragment to facilitate subsequent subcloning of the 5-domain fragment into pDEF2.

5'-GCGTTAAAGCTTCACAGCTCATCACCATGGGCCCTTGGAGCCGCA-3' SEQ ID NO: 14

5'-AGGCGCTCGAGAATCCCCAGAATGGCAGGAATT-3' SEQ ID NO: 15

The E-cadherin cDNA fragment contained in pCR2.1/E-cad#3 was amplified by PCR in a reaction containing 0.5 µl of pCR2.1/E-cad#3, 10 µl of 5X PCR reaction buffer, 1 µl of 10 µM primer Ecad5'Kozak, 1 µl of 10 µM primer E-cad3'(Xho), 1 µl of Advantage™ KlenTaq polymerase mix, and 35.5 µl of H_2O. Amplification conditions included an initial incubation for 1 min at 94°C; 5 cycles at 94°C for 30 sec and 72°C for 4 min; 5 cycles at 94°C for 30 sec and 70°C for 4 min; 25 cycles at 94°C for 30 sec and 68°C for 4 min; and a final 5 min incubation at 72°C.

An aliquot of the PCR reaction was resolved by agarose gel electrophoresis, and a single band of 2.1 kb was observed as expected. The fragment was purified using the Wizard PCR Purification Kit (Promega Corp., Madison, WI), and digested with *Xho*I and *Hin*dIII under standard conditions. The resulting fragment was referred to as 5'-HindIII-Kozak-E-cadherin-XhoI-3'.

The plasmid pDC1/ICAM3.IgG1 was digested with *Xba*I and *Sal*I and a fragment of 908 bp (referred to as 5'-SalI-IgG1-XbaI-3') with a 5' terminal *Sal*I site and a 3' terminal *Xba*I site was purified from a low melting temperature agarose gel (FMC BioProducts, Rockland, ME). This fragment contains the sequences encoding the CH2-CH3 region of human IgG1. The expression vector pDEF2 was linearized in the multiple cloning site with *Hin*dIII and *Xba*I and a three-way ligation reaction was performed which contained the 5'-HindIII-Kozak-E-cadherin-XhoI-3' fragment, linearized pDEF2, and the 5'-SalI-IgG1-XbaI-3' fragment. In this reaction, the 3' *Xho*I site in 5'-HindIII-Kozak-E-cadherin-XhoI-3'was joined in-frame to the 5'-SalI-IgG1-XbaI-3'; both *Xho*I and *Sal*I have compatible 5' overhangs which can be ligated together but cannot be re-digested with either *Xho*I or *Sal*I. An aliquot of the ligation reaction was used to transform the bacterial strain XL-1 Blue (Stratagene, La Jolla, CA). Individual bacterial colonies were grown overnight in LBM containing 100 μg/ml ampicillin, and plasmid DNA was isolated with a Wizard Plus Miniprep Kit (Promega Corp., Madison, WI). The pDEF2/E-cadIgG1 plasmid DNA was digested with *Hin*dIII and *Xba*I and the digestion products resolved by agarose gel electrophoresis. Those clones containing a 2.1 kb fragment were sequenced to ensure that the E-cadherin-IgG1 chimera maintained an open reading frame across the E-cadherin/IgG1 junction.

The pDEF2/E-cadIgG1 clone #3 was found to contain a continuous open reading frame across the E-cadherin/IgG1 junction and was used for CHO cell expression studies described below. The open reading frame of the E-cadherin/IgG1 fusion was not sequenced in its entirety since the DNA fragments contributing to this chimera had been previously sequenced and had not been subjected to PCR amplification.

C. MAdCAM-1-1

1. Isolation of a partial cDNA for human MAdCAM-1-1.

A fragment containing a partial cDNA for MAdCAM-1 was isolated by PCR amplification of Marathon-Ready™ human spleen cDNA library with an Advantage™-GC cDNA PCR Kit (CLONTECH Laboratories, Inc., Palo Alto, CA). Polymerase chain reactions were performed in a Perkin Elmer Cetus (PE Applied Biosystems, Foster City, CA) DNA thermal cycler in a reaction containing 5 µl of Marathon™ human spleen cDNA, 1 µl of 10 µM primer MAdCAM-1 5'#1 (SEQ ID NO:16), 1 µl of 10 µM primer MAdCAM-1 3'#5 (SEQ ID NO: 17), 10 µl of 5.0 M GC-Melt™, 1 µl of 50X dNTP mix, 1 µl of Advantage™ KlenTaq polymerase mix, 10 µl of 5X reaction buffer, and 21 µl of H_2O.

5'-ATGGATTTCGGACTGGCCCTCCTGCT-3' SEQ ID NO: 16
5'-CTCCAAGCCAGGCAGCCTCATCGT-3' SEQ ID NO: 17

Amplification conditions included an initial incubation for 1 min at 94°C; 5 cycles at 94°C for 30 sec and 72°C for 3 min; 5 cycles at 94°C for 30 sec and 70°C for 3 min; 25 cycles at 94°C for 30 sec and 68°C for 3 min; and a final incubation for 5 min at 68°C. An aliquot of the reaction was resolved by agarose gel electrophoresis and a single fragment of 640 bp was detected. The fragment was subcloned into pCR®2.1 and amplified in bacteria using the TA Cloning® Kit (Invitrogen Corp., Carlsbad, CA) following the manufacturer's protocol. Single bacterial colonies were grown overnight in LBM containing 100 µg/ml ampicillin. Plasmid DNA was isolated from the cultures using a Wizard Plus Miniprep Kit (Promega Corp., Madison, WI), and the nucleotide sequence of the subcloned PCR product was determined by DNA sequence analysis. This partial cDNA for MAdCAM-1 begins with the initiation codon and terminates at its 3' end at residue 640 in domain 2. The sequence of this partial MAdCAM-1 cDNA is identical to that previously reported [Shyjan *et al.*, J. Immunol. 156:2851-2857 (1996)].

2. Additional PCR amplification DNA encoding MAdCAM-1 domains 1 and 2

In order to express domains 1 and 2 of MAdCAM-1 as a secreted immunoglobulin fusion protein, it was essential to: (i) restore a Kozak sequence upstream of the initiation codon to allow for efficient protein translation; (ii) add a 5' *Hin*dIII site to facilitate subcloning of the fragment into pDEF2; (ii) extend the open reading frame of the existing partial MAdCAM-1 cDNA to encompass additional amino acid residues needed to encode the entire second domain; and (iv) introduce a *Sal*I site at the 3' terminus of the fragment to facilitate subcloning into pDEF2. These modifications were introduced into the MAdCAM-1 fragment described above by PCR amplification using the primers Mad5'Kozak (SEQ ID NO: 18) and Mad 3' #6 Sal (SEQ ID NO: 19) as shown below.

5'-GCGTTAAAGCTTCACAGCTCATCACCATGGATTTCGGACTGGCCCTCCT-3'

SEQ ID NO: 18

GCTAGTCGACGGGGATGGCCTGGCGGTGGCTGAGCTCCAAGCAGGCAGCCTCATCGT

SEQ ID NO: 19

The PCR reaction included 0.5 µl of pCR®2.1/MAd#4-1 template DNA, 10 µl of 5X PCR buffer, 10 µl of 5.0 M GC Melt™, 1 µl of 50X dNTP mix, 1 µl of 10 µM, 1 µl of 10 µM, 1 µl of 50X Advantage™ KlenTaq polymerase mix, and 25.5 µl of H_2O. The PCR amplification conditions included 94°C, for 1 min; 5 cycles at 94°C for 30 sec and 72°C for 2 min; 5 cycles at 94°C for 30 sec and 70°C for 2 min; 20 cycles at 94°C for 30 sec and 68°C for 2 min; and 68°C for 5 min. An aliquot of the reaction was resolved by agarose gel electrophoresis and a single fragment of ~ 0.7 kb was detected as expected. The PCR product was purified using the Wizard PCR Purification Kit (Promega Corp., Madison, WI) and digested with *Hin*dIII and *Sal*I under standard conditions. The fragment was ligated into *Hin*dIII/*Sal*I digested pBluescript® SK plasmid DNA (Stratagene, La Jolla, CA) under standard conditions, and the sequence of the MAdCAM-1 fragment in pBS-SK/Mad#7 was determined.

3. Generation of MAdCAM-1/Ig Fusion Protein

To generate an expression vector encoding a chimeric domain1/domain2 MAdCAM-1-IgG1 fusion protein, the 702 bp *Hin*dIII-*Sal*I fragment from pBS/Mad#7, the 908 bp *Sal*I-*Xba*I fragment from pDC1/ICAM3.IgG and pDEF2 linearized by digestion with *Hin*dIII and *Xba*I were combined in a ligation reaction. An aliquot from the ligation reaction was used to transform XL-1 Blue bacteria (Stratagene, La Jolla, CA) and the plasmid DNA isolated from single colonies were screened by restriction digestion with *Hin*dIII, *Xba*I, and *Sal*I. One clone, pDEF2/MadIg#1, was found to contain all three fragments and was used to generate stably transfected CHO cell lines as described below.

Example 6

Expression of MAdCAM-1/Ig and E-cadherin/Ig

A. <u>Generation of Stable CHO Cell Lines Expressing MAdCAM-1/Ig and E-cadherin/Ig</u>

For transfection of host CHO DG44 cells with pDEF2/MAdIg or pDEF2/EcadIg, 50 to 100 ug of plasmid was linearized by digestion with the restriction enzyme *Pvu*I. DG44 cells were cultured in DMEM/F-12 medium supplemented with hypoxanthine (0.01 mM final concentration) and thymidine (0.0016 mM final concentration), also referred to as "HT". DG44 cells were prepared for transfection by growing cultures to about 50% or less confluency in treated 150 cm^2 tissue culture polystyrene flasks (Corning Inc., Corning, NY). Cells were collected and resuspended in 0.8 ml of a solution containing HeBS buffer (20 mM Hepes, pH 7.0, 137 mM NaCl, 5 mM KCl, 0.7 mM, Na_2HPO_4 and 6 mM dextrose) with the desired plasmid DNA. The resuspended cells were electroporated at room temperature with a capacitor discharge of 290 V and 960 µF (9 to 11.5 msec pulse). Cells were added to 10 ml DMEM/F-12 supplemented with 5% dialyzed FBS and HT, pelleted by centrifugation, resuspended in 2 ml DMEM/F-12 supplemented with 5% dialyzed FBS and HT ("non-selective media"), and seeded into 75 cm^2 polystyrene tissue culture flasks. After two days growth the cells were

collected and seeded at varying dilutions in DMEM/F-12 supplemented with 5% dialyzed FBS and without HT ("selective media").

Once selection was complete and single cell clones could be identified, a single cell suspension of pooled CHO transfectants was prepared by typsinization. In order to isolate individual clones, the CHO/MAdCAM-1Ig and CHO/E-cadIg transfectants were plated at a density of approximately 1 cell/well in Immulon-4 96-well plates (Dynex Technologies, Inc., Chantilly, VA) under selective conditions. Once single colonies were detected in the 96-well plates, supernatant from each well was screened for the presence of MAdCAM-1/Ig or E-cadherin/Ig fusion protein. Single cell CHO clones producing a human IgG1 protein component were expanded, and those clones producing the greatest level of MAdCAM-1/Ig or E-cadherin/Ig fusion protein were selected for large-scale protein production.

In large-scale protein production, the CHO/MadIg and CHO/E-cadIg clones were expanded in serum-free 5.2 (HT-) media in a spinner flask maintained at 37°C in an atmosphere of 5% CO_2. When cell densities exceeded 10^6 cells/ml., the media was harvested and the spinner flask was provided with fresh 5.2 (HT-) media. The spent media was first centrifuged to remove cell debris, filtered through a 0.22 µm 1 liter filter unit (Corning Inc., Corning, NY), and stored at 4°C.

B. <u>MAdCAM-1/Ig Purification</u>

MAdCAM-1/Ig was purified by affinity chromatography using a protein A-Sepharose® 4 Fast Flow resin column (Flow (Amersham Pharmacia Biotech, Inc., Piscataway, NJ) equilibrated with CMF-PBS. The cell supernatant was cycled through the column at a rate of 4 ml/min. After loading, the column was washed with CMF-PBS until there was no detectable protein present in the eluate. MAdCAM-1/Ig was eluted with 0.1 M acetic acid (pH 3.0) into a tube containing 1M Tris, pH 9.0, and the sample was dialyzed at 4°C against CMF-PBS.

C. <u>Purification of E-cadherin/Ig</u>

E-cadherin/Ig was purified by affinity chromatography using a protein A-Sepharose® 4 Fast Flow resin column (Amersham Pharmacia Biotech, Inc.,

Piscataway, NJ) equilibrated with D-PBS. The supernatant was cycled through the column at a rate of approximately 4 ml/min. After loading, the column was washed with Tris-buffered saline, pH 8.0, containing 1 mM $CaCl_2$ until there was no detectable protein present in the eluate. E-cadherin/Ig was eluted with 0.1 M acetic acid (pH 3.0) containing 1 mM $CaCl_2$ into a tube containing 1M Tris, pH 9.0. Calcium concentration was adjusted to 1 mM and the sample was dialyzed at 4°C against Tris-buffered saline (pH 6.8) containing 1 mM $CaCl_2$.

Example 7
Generation of JY/alpha-E Transfectants

The human B lymphoblastoid cell line, JY, was transfected with the plasmid pMHneo/aE as described above. The transfected population was grown in "selection media" (containing RPMI 1640 media supplemented with 5% FBS, 100 U/ml penicillin G, 100 µg/ml streptomycin sulfate, 2 mM L-glutamine, 1 mM sodium pyruvate, and 1.0 mg/ml G418) and after 14 days, 10^8 G418-resistant JY cells were resuspended in 5 ml of selection media containing 5 µg/ml of the anti-aE monoclonal antibody Ber-ACT8 (DAKO Corp., Carpinteria, CA) and incubated on ice for 1 hour. Cells were collected by centrifugation, and the media was aspirated. The JY/aE transfectants were stained with selection media containing a 1:200 dilution of sheep anti-mouse Ig-FITC (Sigma Corp., St. Louis, MO) on ice for 1 hour. Unbound antibody was removed by centrifugation and the supernatant aspirated. Alpha E-expressing cells were isolated by flow cytometry and subsequently expanded by *in vitro* culture in selection media.

Re-analysis of the sorted JY/aE$^+$ population by flow cytometry with Ber-ACT8 revealed a bimodal population of cells that contained both alpha E-expressing and alpha E-nonexpressing cells. The bimodal population was stained a second time with Ber-ACT8 as previous described, and individual JY/aE cells were sorted into a 96-well Immulon-4 plate containing selection media. Single cell JY/aE$^+$ clones expressing high levels of alpha E were expanded *in vitro* and JY/aE clone #47 was selected for further characterization. This clone, but not the parental JY cells, displayed robust adhesion to recombinant E-cadherin/Ig and the binding was induced with phorbol ester treatment of the cells. Binding of JY/aE clone #47 to

E-cadherin/Ig was blocked by the anti-β_7 integrin antibody FIB504 (ATCC, Rockville, MD), as well as by antibodies to E-cadherin (Zymed Corp., So. San Francisco, CA).

Example 8
Isolation of a JY/alpha D$^+$ Clones

To obtain a JY cell line that stably expresses the $\alpha_d\beta_2$ integrin, JY cells were electroporated with pMHneo/aD as described above and stable transfectants were selected by growth in selection media. After a G418-resistant population of cells had been selected, JY/aD$^+$ cells were stained with the anti-α_d monoclonal antibody 212D and sheep anti-mouse-FITC (Sigma Corp., St. Louis, MO). Single cell JY/aD$^+$ clones were isolated by cell sorting using a flow cytometer as previously described for the isolation of single cell JY/aE$^+$ clones.

Example 9
JY/aE$^+$ Adhesion Assays

A. Compound Dilutions

Adhesion media (350 µl) (RPMI 1640 containing penicillin and streptomycin, L-glutamine, NaPy, and 5% FBS) was aliquotted into each well in rows A, C, E, G of a deepwell 96-well titer plate, 2.0 ml capacity (Beckman Instruments, Inc., Fullerton, CA) in columns 1-11. All compounds to be screened were dissolved in DMSO to a final concentration of 10 mM. Compounds were stored at -20°C, and thawed on the day of use in a 37°C incubator. Each compound (2.1 µl) was pipetted into a single well in columns 3-11, rows A, C, E, and G in the deepwell titer plate. To wells not containing compound (A1 & A2, C1 & C2, E1 & E2, G1 & G2), 2.1 µl DMSO was added. An anti-β_7 monoclonal antibody, FIB504 (ATCC, Rockville, MD), which blocks $\alpha_E\beta_7$ binding activity, was added to wells C2 and G2 at a concentration of 7.5 µg/ml. Each deepwell titer plate was covered to prevent dessication and stored in a 37°C incubator until ready for use.

B. Adhesion Assay

Adhesion assays were performed in 96-well Immunlon 4 plates (Dynex Technologies, Inc., Chantilly, VA) as follows. Each well was coated with 50 µl E-cadherin/Ig (3.0 µg/ml) in D-PBS. Control wells were coated with capture antibody FIB504, to quantitate 100% input cell binding, or coating buffer alone to determine background binding. Following an overnight incubation at 4°C, the plates were washed three times with 200 µl/well D-PBS and blocked with 1% BSA in D-PBS for at least 1 hour. The BSA solution was removed and 100 µl of adhesion media (RPMI 1640 containing penicillin and stretomycin, L-glutamine, sodium pyruvate, 0.1% BSA, and 60 ng/ml PMA), was added to rows B through G, columns 1 through 11.

At this point, 100 µl adhesion media containing a test compound at a concentration of 60 µM, was transferred from the deepwell 96-well titer plate, in triplicate, to the E-cadherin/Ig coated adhesion plate. The outer rows were filled with 300 µl of D-PBS. These plates were transferred to a humidified 37°C incubator with an atmosphere of 5% CO_2.

The adhesion assay was initiated by addition of 100 µl of the JY/aE$^+$ cell suspension to each well of the E-cadherin-coated plate. The final volume in each well was 300 µl adhesion media containing 10^5 cells, PMA (final concentration 20 ng/ml), and the test compound (final concentration 20 µM). The plates were incubated at 37°C for 30 min. Each compound was tested in triplicate.

Adherent cells were fixed by the addition of 50 µl of a 14% glutaraldehyde solution in D-PBS. Plates were washed with water, stained with 100 µl/well 0.5% crystal violet (Sigma Corp., St. Louis, MO) solution for 5 min. Three hundred microliters/well of 70% ethanol was added, and adherent cells were quantitated by determining absorbance at 570 nm. Percentage of cell binding was determined by using the mean values for each triplicate in a given assay in the following formula.

$$\% \text{ binding} = \frac{(\text{A570 (binding to E-cadherin/Ig)} - \text{A570 (binding to BSA)}}{\text{A570 (binding in adhesion media without compound)}} \times 100$$

C. IC50 Determinations

During the initial screening of test compounds, each chemical entity was tested in cell-based adhesion assays at a fixed concentration of 20 μM. Those compounds that blocked JY/$\alpha_E\beta_7$-dependent adhesion to E-cadherin/Ig by 50% or more were subsequently retested at multiple concentrations to determine the inhibitory concentration at which cell binding is reduced by 50%, *i.e.*, the IC50 value.

Of the compounds screened, 40, or 1.4 % of the total library, inhibited $\alpha_E\beta_7$-E-cadherin adhesion by 40% or greater. Approximately 18 of the compounds were identified in the diarylamide library, and 22 compounds were identified in the diaryl sulfide library. Upon re-analysis of these primary hits in IC_{50} determinations, 4 of the 40 compounds were shown to inhibit JY/aE+ binding to E-cadherin with an IC_{50} value of not more than 10 μM. Many of the initial hits were eliminated from further characterization if their initial inhibitory activity was not reproducible; or a compound was shown to inhibit multiple integrin-dependent adhesive events; or the IC_{50} value exceeded 10 μM, or it displayed any cytopathic or cytotoxic effects. The following compounds displayed reproducible inhibitory activity at compound concentrations below 10 μM: Cmpd K, Cmpd W, Cmpd Z, Cmpd D as set out in Table 2. There were several compounds that displayed significant inhibitory activity in the initial screen that failed to inhibit JY/aE+/E-cadherin binding upon re-analysis. It is possible that the activity of some diaryl compounds was lost upon repeated freezing and thawing.

To assess the selectivity of each compound, an IC50 value was determined for additional binding partner compounds JY/$\alpha_V\beta_3$ and vitronectin, JY/$\alpha_4\beta_1$ and VCAM/Ig,, JY/$\alpha_d\beta_2$ and VCAM, JY/$\alpha_L\beta_2$ and ICAM-1, JY/$\alpha_M\beta_2$ and iC3b, and JY/$\alpha_4\beta_7$ and MAdCAM-1.

For each IC50 assay, 50 μl of the ligand diluted in 50 mM bicarbonate buffer (pH 9.6) was dispensed per well of an Immulon-4 plate. A single plate was used to test two different ligands, each in triplicate, The coating concentration for the various ligands was as follows: VCAM-1/Ig at 2.0 μg/ml; ICAM-1/Ig at 5.0 μg/ml; vitronectin at 0.5 μg/ml; MAdCAM-1/Ig at 3.0 μg/ml, and iC3b at 5.0 μg/ml. The capture antibody, *e.g.* anti-CD18 monoclonal antibody TS1/22, was added at a

concentration of 10 μg/ml in 50 μl/well. Ligand-coated plates were covered and stored overnight at 4°C. The following day, the contents of each well was decanted, and each plate was washed three times with 200 μl/well D-PBS. The plate was then blocked by the addition of 300 μl/well of 1% BSA/D-PBS solution. Each plate was again covered and incubated at room temperature for at least 1 hour.

For each IC50 determination, the test compound was serially diluted in DMSO to enable testing at final concentrations of 40 μM, 20 μM, 10 μM, 5.0 μM, 2.5 μM, 1.25 μM, 0.63 μM, 0.32 μM and 0.16 μM. Prior to transfer to the adhesion plate, the compounds were initially diluted by transferring 4.2 μl of the diluted compounds to a 96-well deepwell titer plate containing 0.7 ml/well of RPMI 1640, 0.1% BSA, and 3 ng/ml PMA (Sigma Corp., St. Louis, MO) pre-warmed to 37°C. The 1% BSA/D-PBS blocking solution was decanted from the 96-well Immulon-4 plates and replaced with 0.2 ml of diluted compound. For each 96-well plate to be screened, approximately 8 x 10^6 cells were collected by centrifugation and resuspended in adhesion media (RPMI 1640 containing 0.1% BSA) to a final concentration of 10^6/ml. To prevent PMA-dependent homotypic aggregation in the adhesion assay, the anti-CD18 antibody 22F12C (ICOS Corp., Bothell, WA) was added to the cell suspensions to a final concentration of 10 μg/ml, and the cells were incubated at 37°C for 15 min. This antibody was not added to CD18-dependent adhesion assays involving JY/$\alpha_L\beta_2$, JY/$\alpha_d\beta_2$ or JY/$\alpha_M\beta_2$ and their corresponding ligands ICAM-1, VCAM-1, or iC3b.

The adhesion assay was initiated by addition of 100 μl of the cell suspension to each well of the Immulon-4 plate. The plates were incubated at 37°C for 30 min and adherent cells were fixed for least 1 hour by the addition of 50 μl of a 14% glutaraldehyde solution in D-PBS. The plates were washed with water and stained with 100 μl/well 0.5% crystal violet (Sigma Corp., St. Louis, MO) solution for 5 min. The plates were washed a second time with water to remove excess crystal violet dye, and 300 μl 70% ethanol was added to each well. Adherent cells were quantitated by determining the absorbance at 570 nm in a plate spectrophotometer. The percentage of cell binding was determined by using the mean values for each triplicate in a given assay and the formula below.

$$\% \text{ Binding} = \frac{\text{A570 (binding to ligand) - A570 (binding to BSA)}}{\text{A570 (binding in adhesion media without compound)}} \times 100$$

The four compounds Cmpd K, Cmpd W, Cmpd Z, Cmpd D identified in the primary screen were selected for further specificity profiling, whereby their IC_{50} values were determined in additional integrin-dependent adhesive events. In all cases, the indicator cell line used in the binding assay was treated with 2 ng/ml PMA during the course of the assay to stimulate integrin-dependent adhesion. The IC50 values of these four compounds were determined in adhesion assays as indicated in Table 5.

TABLE 5

Compound	E-cadher $\alpha_E\beta_7$	MAdCAM -1 $\alpha_4\beta_7$	iC3b $\alpha_M\beta_2$	VN $\alpha_V\beta_3$	ICAM-1 $\alpha_L\beta_2$	VCAM $\alpha_4\beta_1$	VCAM $\alpha_d\beta_2$
Cmpd K	3 μM	4 μM	4 μM	6 μM	11 μM	>40 μM	>40 μM
Cmpd D	3 μM	4 μM	7 μM	4 μM	28 μM	>40 μM	>40 μM
Cmpd W	5 μM	5 μM	4 μM	8 μM	30 μM	>40 μM	>40 μM
Cmpd Z	3 μM	11 μM	ND	ND	20 μM	>40 μM	>40 μM

Example 10
Cloning, Expression and Purification of Alpha 1, Alpha 2 and Alpha 11 I Domains

The collagen-binding integrins alpha 1, alpha 2 and alpha 11 contain I domain sequences homologous with the I domain sequences contained in the leukointegrins alpha L, alpha M, alpha X and alpha d. To investigate the possibility that these molecules might be susceptible to modulation through an allosteric regulatory site, the library of test compounds was assessed for the ability to inhibit interactions between these integrins and their ligands collagen and laminin.

The alpha 1 and alpha 2 I domain sequences and alpha 11 were cloned into the bacterial expression vector pET15b (Novagen). Expression of these constructs in *E. coli* results in proteins with an amino terminal histidine tag and the "tagged" protein which can be purified using a nickel column. The cloning of the

alpha 11 was carried out as previously described [Velling, *et al.*, J. Biol. Chem. 274:25735-25742 (1999)].

Both alpha 1 and alpha 2 I domain sequences were cloned into pET15b following PCR amplification to add restriction sites that permit the I domains to be cloned in frame with the histidine tag in the vector. The template for the alpha 1 I domain PCR reaction was a full-length alpha 1 cDNA cloned by hybridization from a spleen cDNA library in vector pcDNA-1 Amp as previously described. The hybridization probe used for this screen was the product of the PCR reaction using the following Alpha1.5 (SEQ ID NO: 20) and Alpha1.3 (SEQ ID NO: 21) primers, respectively:

5'-GACTTTCAGCGGCCCGGTGGAAGACATG-3'	SEQ ID NO: 20
5'-CCAGTTGAGTGCTGCATTCTTGTACAGG-3'	SEQ ID NO: 21

The samples were initially incubated at 94°C for 30 sec followed by 5 cycles of 94°C for 5 sec and 72°C for 2 min; 5 cycles of 94°C for 5 sec and 70°C for 2 min; 25 cycles of 94°C for 5 sec and 68°C for 2 min; and a final incubation of 72°C for 7 min. The PCR products were cloned into the TOPO TA vector pCRII (Invitrogen) and sequenced. The resulting clone was used as a template in PCR using the same conditions as above and the amplification product was gel purified, labeled with ^{32}P using a random primed labeling kit (Boehringer Mannheim), and used as a hybridization probe. Hybridization was performed using ExpressHyb hybridization solution (Clontech) under the same conditions used in the screening for full length alpha 11 cDNA The resulting clone, alpha1/pcdna/111 was used as a template to subclone the alpha 1 I domain.

The alpha1 I domain was amplified by PCR using A1.5Nde (SEQ ID NO: 22) and A1.3Bam (SEQ ID NO: 23) primers, respectively shown below

5'-ATATCATATGGACATAGTCATAGTGCTGG-3'	SEQ ID NO: 22
5'-ATATGGATCCCTAAGACATTTCCATTTCAAATG-3'	SEQ ID NO: 23

The alpha 2 I domain was cloned by PCR using a HUVEC cDNA library in the vector pcDNA-1Amp as template and A2.5Nde (SEQ ID NO: 24) and A2:3Bam (SEQ ID NO: 25) primers, respectively shown below:

5'-ATATCATATGGATGTTGTGGTTGTGTGTG-3'	SEQ ID NO: 24
5'-ATATGGATCCCTATGACATTTCCATCTGAAAG-3'	SEQ ID NO: 25

PCR conditions for amplification of both I domains included an initial incubation at 94°C for 2 min followed by 30 cycles of 94°C for 20 sec; 55°C for 30 sec and 72°C for 45 sec; and a final incubation at 72°C for 7 min. The PCR products were gel purified, digested with *Nde*I and *Bam*HI, gel purified again, and cloned into pET15b previously digested with same enzymes. The resulting clones alpha1/pet/2 and alpha2/pet/27 were sequenced

The alpha 1, alpha 2 and alpha 11 pET15b clones were transformed into the bacterial strain BL21(DE3)pLysS (Stratagene) for expression. Histidine-tagged proteins were isolated from the soluble fraction of the *E. coli* lysate using a Ni –NTA agarose column (QIAGEN) and elution with an imidazole gradient. The eluted proteins were dialyzed against CMF-PBS and biotinylated using EZ-Link Sulfo-NHS-LC-Biotin (Pierce) according to the manufacturer's suggested protocol.

An assay for measuring alpha 1 or alpha 2 I domain binding to collagen in a 96-well plate format involves binding collagen to the wells of a 96-well plate, adding biotinylated alpha 1 or alpha 2 protein to the wells and measuring the amount of collagen bound I domain using europium-coupled streptavidin and time resolved fluorescence. Immulon4 96-well plates were coated with 20 µl/ml of rat type I collagen (Sigma) in CMF-PBS overnight at 4°C. Wells were washed with 250 µl of CMF-PBS two times and blocked with 2.5% BSA in CMF-PBS at 30°C for 1 hr. The wells were washed with 200 µl of CMF-PBS and biotinylated protein was added to the wells at 1 µg/ml in either CMF-PBS with 2 mM $MgCl_2$ and 1% BSA or in TBS with 2 mM $MnCl_2$ and 1% BSA and incubated at 37°C for 3 hours. The wells were washed with 200 µl of the same incubation buffers (without I domain protein) two times and collagen bound biotinylated protein was detected with the addition of 100 µl of a 1:1000 dilution of streptavidin europium (SA-Eu; Wallac) in

SA-Eu dilution buffer (Wallac). Incubation was for 1 hour at room temperature. The wells were washed with 200 µl of incubation buffer six times and 100 µl of Enhancement solution (Wallac; diluted 1:1 with water) was added to each well for 5 minutes at room temperature. Fluorescence was measured using the Eugen program.

Example 11
Identification of Alpha2 Antagonists of Collagen Binding

FACS analysis has indicated that Jurkat cells express both alpha1 and alpha2 integrins. Binding studies using monoclonal antibodies to each of these integrins has shown that Jurkat cell adhesion to rat type I collagen is mediated predominantly through interaction with alpha2. For example, an alpha2 blocking monoclonal antibody has been shown to completely inhibit Jurkat cell binding to type I collagen. In view of this result, Jurkat cells were employed in an adhesion assay as described below to identify inhibitors of alpha2 binding. The assay was carried out using a modification of a procedure previously described [Sadhu, *et al.*, *supra*]

Immulon 4 plates (Dynex Technologies, Chantilly, VA) were coated overnight at 4°C with (i) 50 µl rat type I collagen (Sigma) (20 µg/ml in CMF-PBS), (ii) anti-beta1 monoclonal antibody 3S3 (5µg/ml) in bicarbonate buffer, pH 9.6, (iii) or bicarbonate buffer alone. Plates were washed once with 200 µl/well D-PBS and blocked with 1% BSA (100 µl/well) in D-PBS for 1 hr at room temperature. Wells were rinsed once with 100 µl adhesion buffer containing RPMI and 1% inactivated FBS and 100µl adhesion buffer containing PMA (10 ng/ml final concentration) was added to each well. Adhesion buffer (100 µl) with or without candidate inhibitor (at a final concentration of 20 µM) was added to each well, followed by addition of 100 µl Jurkat cells (1 x 10^6 cells/ml) in adhesion buffer, and incubation carried out at 37°C for 30 min. Adherent cells were fixed by additional of 50 µl/well 14% glutaraldehyde in D-PBS and incubation at room temperature for 2 hr. The plates were washed with dH_2O and stained with 50 µl/well 0.5% crystal violet in 10% ethanol for 5 min at room temperature. The plates were washed in several changes of dH_2O, after which 70% ethanol was added. Adherent cells were quantitated by determining absorbance at 570 nm and 410 nm using a SPECTRmax 250 microplate

spectrophotometer system (Molecular Devices, Sunnyvale CA). The percentage of cell binding was determined using the formula below.

$$\% \text{ Binding} = \frac{A570 - A410(\text{binding to collagen})}{A570 - A410 (\text{binding to mAb 3S3})} \times 100$$

Data was normalized using the formula:

$$\% \text{ of DMSO binding} = \frac{\% \text{ of cell binding, inhibitors}}{\% \text{ of cell binding, DMSO}} \times 100$$

One hundred twenty-one compounds inhibited Jurkat adhesion to rat type I collagen at a level of 50% or greater than the control. IC50 determinations for these inhibitors were assessed in Jurkat adhesion assays as described above except that inhibitors were tested at two-fold dilutions through the concentration range of 0.15 to 40 μM. The IC50 values of 113 of the 121 compounds were determined and 21 of these 121 were selected based on potency in the IC50 range of 2 to 17 μM in the assay and for specificity in showing low level inhibition in the $\alpha_4\beta_7$ binding assay (described herein) and the von Willebrand factor binding assay (described herein). These 21 compounds were further analyzed for specificity and toxicity. In specificity determinations, compounds were tested in a concentration range of 0.15 μM to 20 μM for the ability to inhibit Jurkat cell binding to immobilized VCAM-1/Ig. The assay was carried out in a manner similar to the collagen adhesion assay described above except that cells were coated with VCAM-1/Ig instead of collagen. Binding to VCAM-1 was dependent on surface expression of $\alpha_4\beta_1$. These results are shown in Table 6.

For 21 compounds, toxicity of Jurkat cells was assessed following a four hr or 24 hr incubation. LD50 concentrations were determined using a CellTiter 96® Aqueous One Solution Cell Proliferation Assay System (Promega) according to the manufacturer's suggested protocol. A two-fold serial dilution series of each compound was tested in a concentration range of 40 μM to 0.15 μM. Results from the toxicity assay are shown in Table 6.

TABLE 6

COMPOUND	$\alpha_2\beta_1$/COLLAGEN EC50 (μM)	$\alpha_7\beta_1$/VCAM EC50 (μM)	TOXICITY LD50 (μM)
Cmpd AD	2	>20	3
Cmpd T	4	15	25
Cmpd AF	6	>20	35
Cmpd AI	6	>20	33
Cmpd AG	7	>20	35
Cmpd AE	7	>20	30
Cmpd Y	7	>20	25
Cmpd J	8	>20	>40
Cpmd X	8	>20	30
Cmpd M	8	>20	25
Cmpd AL	8	>20	20
Cmpd AJ	8	>20	38
Cmpd AK	9	>20	35
Cmpd AH	9	>20	38
Cmpd AB	13	>20	>40
Cmpd A	14	17	>40
Cmpd U	15	>20	>40
Cmpd G	16	>20	>40
Cmpd E	16	>20	>40
Cmpd B	17	>20	>40
Cmpd AN	17	>20	>40

Example 12
Identification of Alpha1 Antagonists of Collagen Binding

Chinese hamster ovary (CHO) cells do not express endogenous collagen receptors. Accordingly, CHO cells were transfected with a full-length alpha1 expression construct, alpha1/pDC-1/1. The full-length alpha1 insert was

removed from a clone in the vector pLEN [Briesewitz *et al.*, JBC 268:2989-2996 (1993)] and subcloned into the pDC-1 to generate the clone alpha1/pDC-1/1. Transfectants were grown in selective media (DMEM/F12 with 10% FBS) and cloned by limiting dilution. Alpha1 expressing clones were identified by staining the cells with a blocking alpha1 monoclonal antibody (antibody 5E8D9; Upstate Biotech) and determining expression levels by FACS analysis. These cells were demonstrated to adhere to type IV collagen in an alpha1-dependent manner using the blocking alpha1 monoclonal antibody (Upstate Biotech; clone 5E8D9) which was shown to inhibit this adhesion. In view of this result, the alpha1 transfected CHO cells were used in an adhesion assay as described below to identify inhibitors of alpha1 binding. This assay is a modification of the procedure used to identify alpha2 antagonists described above.

Immulon 4 plates were coated overnight at 4°C with either (i) 50 µl per well human type IV collagen (Sigma) (0.5 µg/ml in CMF-PBS), (ii) the anti-alpha1 monoclonal antibody 5E8D9 in bicarbonate buffer, pH 9.6, or (iii) bicarbonate buffer alone. Plates were washed twice with D-PBS and blocked with 1% BSA (100 µl/well) in D-PBS for 1 hour at room temperature. Wells were rinsed once with 100 µl/well adhesion buffer (DMEM/F12 media with no serum). Adhesion buffer (200 µl) with or without candidate inhibitor was added to each well followed by the addition of 100 µl of alpha1 transfected CHO cells in adhesion buffer. CHO cells were previously recovered using versene and rinsed 3 times in DMEM/F12 media containing 10% FBS. Cells were resuspended in adhesion buffer at a density of 0.75 x 10^6 cells/ml. Incubation of the alpha1-transfected CHO cells on type IV collagen was carried out at 37°C for 30 minutes. Adherent cells were fixed by additional 50 µl/well 14% glutaraldehyde in D-PBS and incubation at room temperature for 2 hours. The plates were washed with dH2O and stained with 50 µl/well 0.5% crystal violet in 10% ethanol for 5 minutes at room temperature. The plates were washed in several changes of dH2O after which 70% ethanol was added. Adherent cells were quantitated by determining absorbance at 570 nm and 410 nm using a SPECTRmax 250 microplate spectrophotometer system (Molecular Devices,

Sunnyvale CA). The percentage of cell binding was determined using the formula below.

$$\% \text{ binding} = \frac{(A570\text{-}A410(\text{binding to collagen})}{(A570\text{-}A410(\text{binding to mAb 5E8D9})} \times 100$$

Data was normalized using the formula:

$$\% \text{ of DMSO binding} = \frac{\% \text{ of cell binding, inhibitors}}{\% \text{ cell binding, DMSO}} \times 100$$

Sixty-four compounds inhibited alpha1-transfected CHO cell adhesion to type IV collagen by a level of 50% or greater than the DMSO-control. EC50 determinations for these compounds were determined in alpha1-transfected CHO cell adhesion assays as described above, except that the inhibitors were tested at two-fold dilutions through the concentration range of 0.15 μM to 20 μM (*i.e.*, 0.15 μM, 0.3125 μM, 0.625 μM, 1.25 μM, 2.50 μM, 5 μM, 10 μM, 20 μM). The EC50 values for these compounds ranged from 0.5 μM to 18 μM. These compounds were further analyzed for selectivity and toxicity.

For initial specificity testing, the compounds were tested in a concentration range of 0.15 μM to 20 μM for the ability to inhibit alpha2-transfected CHO cell adhesion to type I collagen. For this assay CHO cells were transfected with an alpha2 expression construct, alpha2/pDC-1/8. The original alpha2 construct was in the expression vector pcDNA-3 and was a Genestorm clone purchased from Invitrogen. The alpha2 sequence was subcloned into pDC-1 resulting in the clone alpha2/pDC-1/8. Alpha2-expressing cells were cloned and analyzed by FACS using an alpha2 monoclonal antibody, A2-IIE10 (Upstate Biotech). A CHO cell line expressing moderate levels of alpha2 was identified and used in adhesion assays as described above for alpha1. The only differences in the alpha2 adhesion assay included (i) using immobilized rat type I collagen (Sigma) in place of the type IV collagen and (ii) using the alpha2 monoclonal antibody, A2-IIE10, in place of the alpha1 monoclonal antibody. Most compounds had a narrow range of specificity for

alpha1 compared with alpha2. These compounds were about 1-3 fold more potent in inhibiting alpha1 dependent adhesion than for inhibiting alpha2 dependent adhesion.

The toxicity of the compounds was assessed in a 4 hour assay using the alpha1-transfected CHO cells. LD50 concentrations, (or "Lethal Dose 50"), as used herein, is the compound concentration necessary to kill 50% of the cells over a defined time interval. LD50 concentrations were determined using a CellTiter 96 Aqueous One Solution Cell Proliferation Assay System (Promega) according to the manufacturer's suggested protocol. A two-fold serial dilution series of each compound was tested in a concentration range of 40 μM to 0.15 μM. Toxicities for these compounds ranged from 2.5 μM to 40 μM.

Thirty-two compounds which were chosen based on their potency, selectivity and toxicity profiles were further analyzed for specificity. Compounds were tested for inhibiting adhesion of more distantly related I domain-containing integrins alphaL (LFA-1) and alphaM (Mac-1). For alphaL, the compounds were tested for inhibition of JY8 cell adhesion to ICAM-1.

ICAM-1/JY-8 Cell Adhesion Assay

Biologically relevant activity of the compounds in the present invention was confirmed using a cell-based adhesion assay that measures the ability of the compounds to block adherence of JY-8 cells (a human EBV-transformed B cell line expressing LFA-1 on its surface) to immobilized ICAM-1, as follows. Compounds were screened for the inhibition of LFA-1 dependent adhesion, as described with respect to the alpha1 assay, with some modifications. Plates were coated with ICAM-1 Ig protein (5 μg/ml in sodium bicarbonate buffer solution) instead of type IV collagen. JY cells were used in place of K562 [α_1] cells. The capture monoclonal antibody used was 22F12C (at 5 μg/ml in sodium bicarbonate buffer solution) in place of an alpha1 monoclonal antibody.

For alphaM, the compounds were tested for inhibition of Mac-1 transfected JY cell adhesion to iC3b (assay described in Example 2). For both assays, compounds were tested in a 2-fold dilution series in a concentration range from 20 μM to 0.15 μM. Most compounds were 1-10 fold more effective at

inhibiting alpha1 dependent adhesion than inhibiting LFA-1 and MAC-1 dependent adhesion. These compounds were also analyzed for toxicity in a 4 hour assay with the JY cells as described above for the CHO cells.

A second alpha1-dependent cell adhesion assay was developed to further assess the alpha1 antagonists identified. K562 cells, a myeloid leukemia cell line, was transfected with a full-length alpha1 expression construct alpha1/pMHneo/40. The alpha1/pMHneo/40 construct was generated by subcloning the full length alpha1 sequence into the expression vector pMH-neo [Hahn *et al.*, Gene 127:267-268 (1993)]. Transfectants were selected with 0.5 mg/ml G418. In order to further select for alpha1-expressing cells, the transfectants were panned for adhesion to type IV collagen. For the panning, tissue culture plates were coated with 20 mg/ml of human type IV collagen (Sigma) in CMF-PBS for 1 hour at 37°C. The plates were washed with binding buffer (RPMI with 10% FBS) and the alpha1-transfected K562 cells were added in binding buffer containing 20 ng/ml PMA. After incubation at 37°C for 1 hour, the plates were washed to remove unbound cells. Adherent cells were removed with versene and diluted with binding buffer. After panning, K562 cell lines expressing alpha1 were obtained and used for further screening described below.

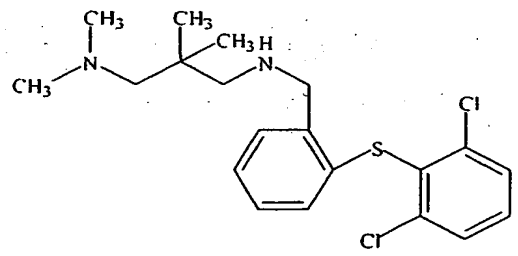
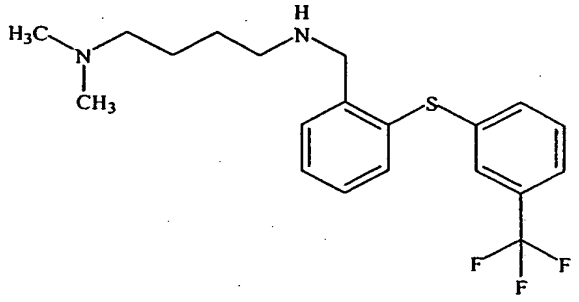
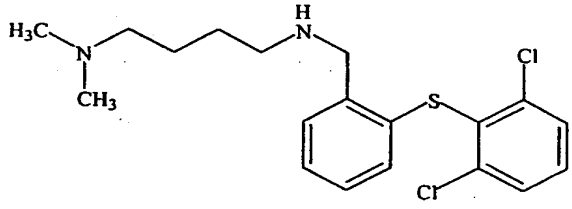
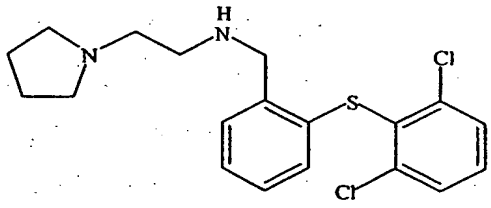
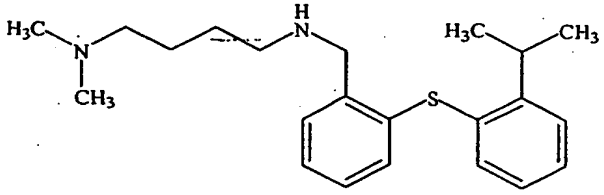
Twenty-one alpha1 antagonists identified in the CHO cell adhesion assay were further analyzed in an alpha1-transfected K562 cell adhesion assay. The cell adhesion assay was performed as described above for the CHO cell assay except that RPMI was used as the adhesion buffer. The potencies of the alpha1 antagonists were similar in the K562 and CHO cell adhesion assays with most EC50 values for most compounds falling within a 1-3 fold range between the two assays. The compounds were also analyzed for toxicity with the K562 cells in a 4 hour assay using the CellTiter 96 Aqueous One Solution Cell Proliferation Assay System described above. The toxicities (LD50) of most compounds was similar in the K562 and the CHO assays. The LD50 values between the two assays varied by less than 2-fold for the majority of compounds.

The structures of five alpha1 antagonists are shown in Table 7. These compounds have EC50 values in the range of 0.5 - 1.5μM. These compounds have

narrow specificity for alpha1 over alpha2 (1 - 4 fold), and greater selectivity over more distantly related integrins, such as LFA-1 and Mac-1 (3 - 10 fold). The window between potency (EC50) and toxicity (LD50) ranges from 6 - 20 fold.

5

TABLE 7

AT	
AU	
AV	
AW	
AX	

Example 13
Expression and Purification of Alpha1 I domain and its Usage in a Biochemical Assay

An alpha1 I domain construct was generated for expressing the alpha1 I domain as a histidine tagged protein in *E. coli*. The histidine-tagged protein was used in co-crystallization experiments to determine the 3-dimensional structure of the alpha1 I domain complexed with inhibitors. The histidine tagged protein was also used to assess alpha1 antagonists in a biochemical assay by measuring the binding of the alpha1 I domain to immobilized collagen.

The alpha1 I domain was cloned as follows. A polynucleotide encoding the alpha1 I domain was PCR amplified using the A1.I.Bam (SEQ ID NO: 26) and A1.I.Pst (SEQ ID NO: 27) primers shown below and the vector alpha1/pDC-1/1 as template.

A1.I.Bam : CGGATCCCCCACATTTCAAGTCGTGAAT SEQ ID NO: 26
A1.I.Pst : GCTGCAGTCATATTCTTTCTCCCAGAGTTTT SEQ ID NO: 27

PCR conditions included an initial incubation at 94°C for 2 minutes, followed by 30 cycles of 94°C for 30 seconds; 55°C for 30 seconds and 72°C for 30 seconds; and a final incubation of 72°C for 7 minutes. The resulting PCR product was gel purified, digested with BamHI and PstI, gel purified again and then cloned into the vector pQE30 (Qiagen) previously digested with BamHI and PstI. The resulting clone alpha1/pQE30/2 was verified by sequencing.

The alpha1/pQE30/2 construct was transformed into *E. coli* strain M15(pREP4) (Qiagen) for protein expression. Histidine-tagged alpha1 I domain was solubilized from purified inclusion bodies using 6 M guanidine and then snap refolded by dilution in buffer without guanidine. The solubilized alpha1 I domain was purified using a Ni-NTA agarose column (Qiagen) and elution with an imidazole gradient.

The purified alpha1 I domain was used in direct binding assays with immobilized type IV collagen as follows. Costar Immulon4 plates (96 well) were coated overnight with either (i) 50 µl/well of human collagen IV protein (Sigma) at

40 μg/ml in CMF-PBS, (ii) anti-alpha 1 I-domain monoclonal antibody (Immune Diagnostics) at 10 μg/ml in CMF-PBS (positive control), or (iii) CMF-PBS alone (negative control). Plates were incubated overnight at 4°C. The next day, media was removed and the plates were blotted dry, after which 150 μl/well of 2% BSA in CMF-PBS containing 0.05% Tween-20 was added to block the plates, and plates were incubated further at 37°C for 1 hour. Media was again removed from the plates which were blotted dry, and then washed twice with 150 μl/well of CMF-PBS containing 0.05% Tween-20 and 5 mM $MgCl_2$ (PBS/T/Mg). Approximately 50 μl/well of PBS/T/Mg containing 2X compound, DMSO, anti-alpha 1 I-domain monoclonal antibody, isotype-matched control monoclonal antibody or no inhibitor was added to the plates, after which 50 μl/well of PBS/T/Mg containing 20 μg/ml of purified alpha 1 I-domain was added and plates were incubated for 30 minutes at 37°C. Media was removed and the plates were blotted dry, then washed twice with 100 μl/well PBS/T/Mg, after which 100 μl/well PBS/T/Mg containing 1 μg/ml anti-penta-His monoclonal antibody (Qiagen) was added. The plates were then incubated for 30 minutes at 37°C, the media was removed, and the plates blotted dry. The plates were then washed twice with 100 μl/well PBS/T/Mg, 100 μl/well PBS/T/Mg containing a 1:20,000 dilution of GAM-HRP (Sigma) was added, and the plates were incubated for 30 minutes at 37°C. Media was removed, and the plates were blotted dry. The plates were washed twice with 100 μl/well PBS/T/Mg and 100 μl/well of Substrate Buffer containing 150 μl/l of H_2O_2 and a 1:100 dilution of TMB substrate stock was added to each well and the plates were developed in the dark for 30 minutes at room temperature. Fifty μl/well of 15% H_2SO_4 was then added to stop the reaction and the plates were analyzed by $A_{450} - A_{670}$ on a spectrophotometer. The specific signal was determined by subtracting background binding to "negative control" wells.

An additional assay was developed using Europium-labeled alpha 1 I domain and bound I domain was directly detected after washing using time resolved fluorescence (TRF). In this assay, purified alpha-1 I domain was labeled with Europium using a DELFIA Europium-labeling kit according to the manufacturer's suggested protocol (Wallac). Costar Immulon4 plates (96 well) were coated with

100 µl of 25 µg/ml of human collagen IV protein (Sigma) in CMF-PBS/1 µM $MgCl_2$, and incubated overnight at 4°C. Plates were then washed 3 times with TBS/T (20 mM Tris, pH 8.0; 150 mM NaCl; 0.02% Tween-20) and 1mM $MgCl_2$ (TBS/T/Mg), 200 µl per well. Plates were then blocked with CMF-PBS/1mM $MgCl_2$/2% BSA, 100µl per well for 1 hour at 37°C. Plates were washed again, then probed with 5 g/ml Europium I domain in RPMI/5%TBS/1mM $MgCl_2$, 100µl per well, and incubated at 37°C for one hour. Plates were then washed again and developed by adding 100 µl/ well Enhance (Wallac) and analyzed on a Victor plate reader by time resolved fluorescence.

Example 14
Expression and Purification of Alpha1Beta1 Leucine Zipper Protein and its Usage in a Biochemical Assay

In order to develop a more physiologically accurate biochemical assay, an alpha1beta1 leucine zipper protein was generated. Expression constructs were prepared individually encoding the full length extracellular domains of alpha1 and beta1 without the transmembrane and cytoplasmic tail polypeptide sequences. Removal of the transmembrane regions allows these proteins to be secreted from transfected cells providing easy purification. The transmembrane sequences were replaced with the acidic and basic leucine zipper sequences respectively. See generally, Chang *et al.*, PNAS 91:11408-11412 (1994).

The extracellular domain of alpha1 was subcloned from the original alpha1 clone in pLEN [Briesewitz *et al.*, JBC 268:2989-2996 (1993)]. The extracellular domain of beta1 were subcloned from the full length beta1 clone He6.1.2/pcDNA-1Amp. This clone was obtained by screening a Hela cDNA library by hybridization. The leucine zipper sequences promote the formation of the alpha1beta1 heterodimer. These constructs were generated using the same leucine zipper sequences and vectors described in U.S. Patent No. 6,251,395, issued on June 26, 2001, Example 14 of which is hereby incorporated herein by reference for its description of methods for constructing leucine zipper proteins. The alpha1 and beta1 leucine zipper constructs were co-transfected into CHO cells which were then maintained in DMEM/F12 media with 10% dialyzed FBS. Supernatant was

collected and the secreted alpha1beta1 heterodimer was purified using chromatography over CNBr-activated Sepharose 4B (Pharmacia) coupled with an anti-leucine zipper monolconal antibody which recognizes both chains of the leucine zipper. For use in biochemical assays, purified alpha1beta1 leucine zipper protein was Europium labeled using a DELFIA Europium-labeling kit according to the manufacturer's suggested protocol. Binding of the labeled alpha1beta1 leucine zipper protein to immobilized collagen was measured by time resolved fluorescence. The heterodimer assay was set up essentially the same as the Europium labeled I domain assay, with the exception that Europium labeled heterodimer in CMF-PBS/1mM $MgCl_2$/2% BSA was substituted as the probe for the Europium labeled I domain.

Example 15
Von Willebrand Factor/gpIb-CHO Static Cell Adhesion Assay

The A11 domain in von Willebrand factor (vWf) is homologous to I domains found in other proteins. To investigate the possibility that these molecules might be susceptible to similar modulation as described above, the library of test compounds were tested for the ability to modulate vWf binding to gpIb.

Round-bottom (RB) glass plates were coated overnight at 4°C with 50 µl of 1 µg/ml bovine vWf (bvWf) in CMF-PBS. Control wells include wells that were coated with 5 µg/ml of vWf at 50 µl/well, or with fibrinogen at 10 µg/ml. The next morning, the plate was washed once with 200 µl of CMF-PBS, blocked with 200 µl of 2.5% gelatin for 1 hr at 37°C, and washed three times with 200 µl CMF-PBS.

CHO cells transfected with DNA encoding glycoprotein(GP) Ib-IX [Cranmer, *J. Biol. Chem. 274*:6097-6106(1999)] were grown in DMEM/F12 with 10% FCS, antibiotics, glutamine and 5-hydroxytryptophan supplemented with 400 µg/ml G418 and 200 µg/ml zeocin (Invitrogen). Confluent cells were washed once with CMF-PBS and incubated with warm Versene in incubator for 5 min. Cells were collected and resuspended in Tyrode's solution (Sigma) with 4 mM EDTA at a density of 2 x 10^6 cells/ml.

The library of test compounds were diluted in Tyrode's/EDTA to 20 μM and 50 μl of the diluted compound was added to each well to a final concentration of 10 μM. For the control, 1 μl 100%DMSO was added to 300 μl cell, with the final concentration of DMSO 0.3%. In a control with a known vWf inhibitor, aurin-tricarboxylic acid (ATA, Sigma) was dissolved in 100% DMSO to 20 mM, diluted with Tyrode's /EDTA to 20 μM, and 50 μl/well to final concentration of 10 μM was added.

Cells were added to each well at a density of 10^5 cells/well in 50 μl and the plates rocked for 40 min at room temperature. The non-adherent cells were removed by aspiration, 200 μl CMF-PBS was added, the plates vortexed and the buffer removed. Calcein was added (50 μl/well of a 2 μM stock) and the plates incubated at room temperature for 1 hr to label adherent cells. Fluorescence was measured on a Millipore CytoFluor 2350 fluorimeter to quantitate adherent cells. A number of compounds having IC50 values less than 20 μM were identified.

Example 16
CD11b-Mediated Neutrophil Adhesion to Fibrinogen

The adhesion assay described above for CD18/CD11b- (Mac-1)-mediated adhesion of HL-60 cells to ICAM-1 was carried out with the following modifications. Each well was coated overnight at 4°C with 50 μl of glycophorin (10 μg/ml), fibrinogen (5 μg/ml) or with anti-CD18 monoclonal antibody (22F12C, 5 μg/ml) and anti-CD11b monoclonal antibody (44AACB, 5 μg/ml) in 50 mM bicarbonate buffer (pH 9.6). Plates were blocked with 1% human serum albumin and no blocking antibody was used. Neutrophils were isolated from fresh heparin whole human blood by density gradient centrifugation and 100 μl of the cells (4 x 10^6 cells/ml) in adhesion buffer was added to each well. Plates were incubated at 37°C for 10 minutes.

Over 1000 compounds were screened, and several had IC50 values ranging from 1 μM to 40 μM. Nine compounds were found to have IC50 values below 10 μM [Cmpd S, Cmpd R, Cmpd N, Cmpd O, Cmpd P, Cmpd Q, Cmpd L, Cmpd V, and Cmpd F, as set out in Table 2. After SAR efforts based on compound Cmpd S (which initially showed an IC50 of 1 μM), inhibition potency for

compounds improved to less than 200 nM (for Cmpd AA and Cmpd AC) and several compounds showed complete inhibition at 20 μM [compounds ranging from Cmpd Z to Cmpd AM]. With the exception of Cmpd Z all compounds selected for their ability to inhibit $\alpha_E\beta_7$/E-cadherin also antagonize the other β_7 integrin, $\alpha_4\beta_7$. These compounds also exhibited minimal inhibitory activity towards $\alpha_L\beta_2$, $\alpha_d\beta_2$, and $\alpha_4\beta_1$. Also, relative to $\alpha_E\beta_7$/E-cadherin, these compounds display limited selectivity (less than 2-fold) for $\alpha_M\beta_2$ and $\alpha_V\beta_3$.

Example 17
Development of Inhibitors of Rac1 Guanine Nucleotide Exchange Reaction

Rac proteins are not active when bound to GDP, but are activated by the exchange of GDP for GTP. The exchange of GDP for GTP in Rac proteins is catalyzed by guanine nucleotide exchange factors (GEFs) such as, Vav1 and Tiam1 [Aghazdeh *et al.*, Cell 102:625 (2000); Worthylake *et al.*, Nature 408:682 (2000)]. Due to the importance of Rac proteins in the control of cell proliferation, antagonists of the Rac guanine nucleotide exchange reaction and, in particular, small molecules that interfere with the exchange of GDP for GTP of Rac1 in the presence of Tiam1, are of considerable interest for the methods and compositions of the present invention.

A. Cloning and expression of Rac1 and Tiam1:

Rac1 and the DH-PH domain of Tiam1 were cloned using standard recombinant DNA procedures [Disbury *et al.*, J. Biol. Chem. 264:16378 (1989)]. Rac1 was expressed in *E. coli* as a GST fusion protein using the vector pGEX2T in accordance with previously described methods [Self and Hall, Meth. Enzymol. 256:3 (1995)]. Purified thrombin-cleaved Rac1 protein was used in the assay.

The Tiam1 DH-PH domain expressed as a fusion protein containing a carboxy terminal 6XHis tag using the plasmid pET28a described by Rossman and Campbell, Meth. Enzymol. 325:25 (2000).

B. Guanine nucleotide exchange assay:

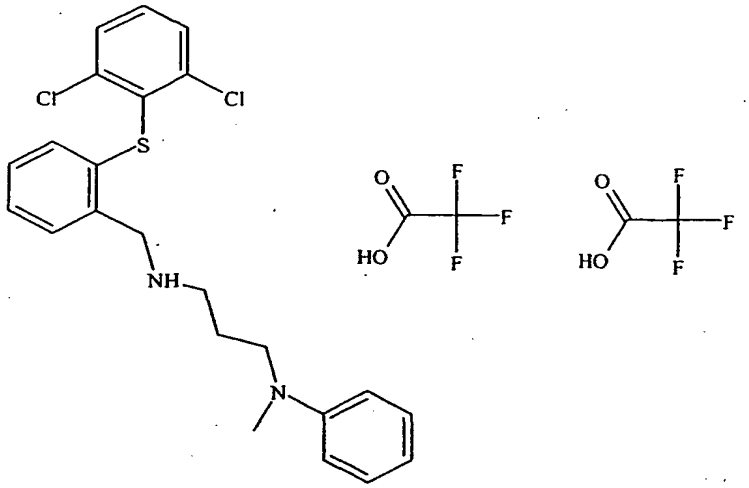
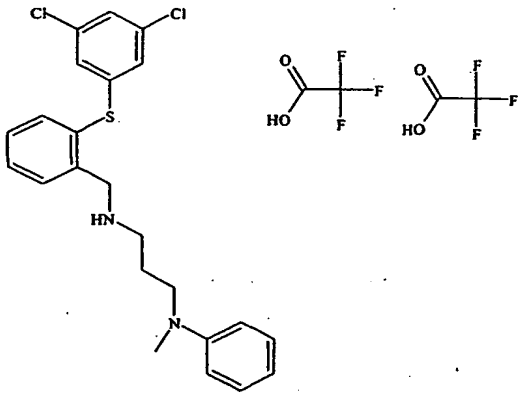
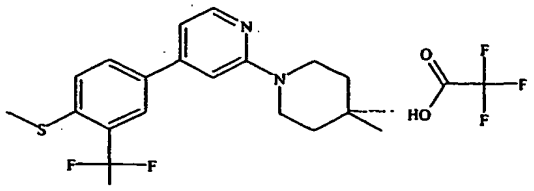
The Tiam1-catalyzed exchange of GDP for GTP of Rac1 was carried out essentially according to the procedure described by Crompton *et al.*, J. Biol.Chem. 275(33):25751 (2000). GDP-bound Rac1 was incubated with [α^{32}P]-labeled GTP, in the presence of Tiam1 and nucleotide exchange was monitored by following the increase in radioactivity bound to Rac1. Free radioactivity was removed by placing the reaction mixture in the well of a 96-well plate and filtering out the fraction of [α^{32}P]GTP that is not bound to Rac1. During the screen for the nucleotide exchange antagonists, the compounds were used at 10 μM concentration. The compounds analyzed by the screening methods are further described below.

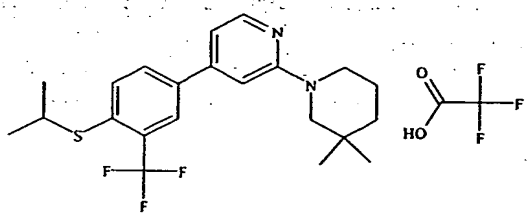
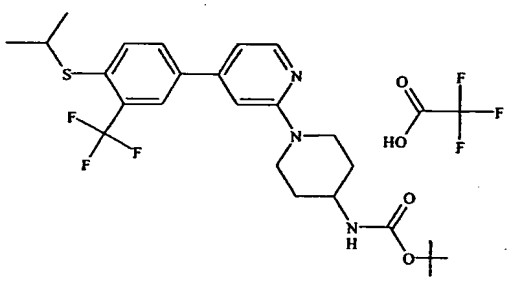
C. Cell proliferation assay:

Rat embryonic fibroblast (REF) and Jurkat cells were selected as representatives of fibroblastic and T cells respectively in order to test the effect of Rac1 guanine nucleotide exchange inhibitors on cell proliferation. REF cultured in the medium RPMI ("REF-R cell culture") was obtained as described by Nobes, Meth. Enzymol. 325:441 (2000). REF-R or Jurkat cells in complete RPMI and 10% fetal bovine serum (FBS) were plated into 96 well plates in duplicate, 10 x 10^3 cells/well. After 21 hrs (for Jurkat), and 45 hrs (for REF-R), AlamarBlue (Serotec) was added, and cells were returned to the incubator (37°C, 5% CO_2) for additional 3 hrs. Results were obtained with SpectraMaxGEMINI (Molecular Devices).

Compounds that inhibit the Rac1 guanine nucleotide exchange reaction by at least 50% of the control were obtained. The IC50 value of several of the compounds were determined for the guanine nucleotide exchange reaction of Rac1, in the presence of Tiam1. The five structures shown in Table 8 represent the most potent inhibitors, and IC50 values for the guanine nucleotide exchange reaction for these compounds are also included therein.

TABLE 8

Compound	IC50 (μM)
AY 	1.9
AZ 	3.1
AAA 	3.5

AAB 	4.7
AAC 	5

Example 18
Inhibition of Bacterial Proteins

Three microbial enzymes containing Rossmann fold structures were identified as candidates for screening with the library of test compounds. Selection was based on (i) presence of the Rossmann structure; (ii) expression patterns in prokaryotic and eukaryotic cells; (iii) clinical importance; and (iv) functional importance to bacterial growth and survival. Two of the selected proteins, dihydrodipicolinate reductase (DHPR or DapB) and enoyl-acp reductase (ENR), catalyze electron transfer from NADH to a substrate and are integral to biosynthetic pathways for lysine synthesis and fatty acid synthesis, respectively. The third and fourth proteins, *E. coli ras*-like GTPase (ERA-GTAse) and yihA (also a GTPase), are involved in translation and cell cycle regulation.

Modulation of DapB activity is assessed using an optical assay that involves synthesis of dihydrodipicolate from aspartate semialdehyde. The assay utilizes dihydrodipicolate synthease (DapA) to first synthesize dihydrodipicolinate, followed by addition of NADH and DapB. A coupled reaction is necessary because dihydrodipicolate is an unstable compound. The change in absorbance in the presence and absence of a test compound resulting from NADH conversion to NAD^+ is monitored at 340 nm.

Identification of modulators of ENR is carried out in a similar manner, but in a single step reaction. Briefly, NADH and ENR are first incubated, followed by addition of substrate, (either crotonyl-CoA or crotonyl-ACP). Again, the change in absorbance in the presence and absence of a test compound resulting from NADH conversion to NAD^+ is monitored at 340 nm.

In view of the fact that optical assays require large amount of substrate, *i.e.*, crotonyl CoA, and the fact that several test compounds absorb at the same wavelength as NADH, alternative thin layer chromatography (TLC) and plate-based assays were designed to identify modulators of ENR using radiolabeled NADH.

The TCL method measures conversion of ^{32}P-NADH to NAD^+ in the presence of lithium chloride which causes the two sates to separate on PEI membranes after a 5 to 10 min run time. Radiolabeled spots are measured on a

Storm Phosphoimager and the ratio of NAD^- to NADH is calculated. An increase in the ratio of NADH to NAD^+ in the presence of a test compound is indicative of inhibition of the conversion. The control reaction is optimized to measure conversion in the linear range. Practical application of this assay was demonstrated using a commercially available enzyme inhibitor. The TLC method is particularly useful for small scale screening.

For large scale screening, the plate based assay is designed to utilize the same reagents. This assay exploits the charge difference between NADH and NAD^+ to permit separation. Positively-charged DEAE-cellulose membrane is used to selectively trap ^{32}P-NADH which has a net negative charge greater than NAD^+. Trapped NADH is detected using scintillation counting and increased signal in the presence of a test compound indicates enzyme inhibition.

For ERA-GTPase, a one step assay is carried out to identify modulators. The transfer of labeled phosphorus in the conversion of GTP to GDP is measured in the presence and absence of a test compound, the label being detected in a scintillation counter using an assay routinely practiced in the art. Conditions for the ERA GTPase assay can also be utilized in screening for yihA modulators.

Example 19
Identification of HPPK Antagonists

The enzyme 6-hydroxymethyl-7,8-dihydropterin pyrophosphokinase (HPPK) is part of the *de novo* folate biosynthetic cascade and catalyzes the transfer of pyrophosphate from ATP to 6-hydroxy-7,8-dihydropterin (HMDP) [Richey *et al.*, *J. Biol. Chem.* 244:1582-1592 (1969)]. HPPK is expressed in both gram positive and gram negative bacteria, fungi, and protozoa, but not in higher eukaryotes. Accordingly, HPPK represents a novel target for the development of antibiotics with anti-folate activity.

1. Isolation of the *E. coli* HPPK Gene

The *E. coli* HPPK gene was isolated by PCR amplification of *E. coli* genomic DNA with the following oligonucleotide primers specific for the 5' (SEQ ID NO: 28) and 3' (SEQ ID NO: 29) ends of the HPPK gene:

5'EcHPPK	5'-GTAGATGACAGTGGCGTATATT-3'	SEQ ID NO: 28
3'EcHPPK	5'-GCCTTACCATTTGTTTAATTTGT-3'	SEQ ID NO: 29

PCR was performed in a Perkin Elmer Cetus (PE Applied Biosystems, Foster City, CA) DNA thermal cycler under standard conditions. *See* generally, Ausubel *et. al*, Current Protocols in Molecular Biology, Vol. 3, p. 15.1.1 - p.15.1.15 (1999). The amplification products were then analyzed by agarose gel electrophoresis to determine the approximate size of the PCR product, and a single DNA fragment of approximately 487 bp was detected, as anticipated.

The HPPK PCR product was ligated to the vector pCRII-TOPO (Invitrogen Corp., Carlsbad, CA) according to the manufacturer's protocols. *E. coli* strain TOP10 (Invitrogen Corp., Carlsbad, CA) was transformed with an aliquot of the ligation reaction, as recommended by the manufacturer, and single bacterial colonies were isolated and grown overnight in LBM media containing 100 µg/ml carbenicillin.

Plasmid DNA was isolated from 2 ml cultures of the single colonies using the Wizard Plus Miniprep Kit (Promega Corp., Madison, WI). The DNA sequence of the *E. coli* HPPK PCR product in the plasmid pCRII-TOPO/EcHPPK was determined to be correct, having the amino acid sequence set out in SEQ ID NO: 30.

2. Generation of His(6)-HPPK Expression Constructs

In order to facilitate the purification and detection of *E. coli* HPPK, the following changes were made to the *E. coli* HPPK coding sequence during a subsequent PCR amplification: 1.) the amino terminus of HPPK was modified to incorporate an additional 6 histidine residues, and 2.) unique restriction sites were added to the 5' and 3' ends of the coding region to facilitate subcloning of the PCR fragment into the expression vector pBAR5. Methods for the subcloning of a similar PCR fragment into an expression vector have been previously described in U.S. Patent No. 5,847,088, issued December 8, 1998, Example 8 of which is hereby incorporated herein by reference. The 5' PCR primer included an NcoI restriction

site followed by sequences encoding the additional amino acid residues "MGHHHHHHGG" (SEQ ID NO. 31) as shown below:

5'EcHisHPPK SEQ ID NO: 32
5'-CGCCATGGGCCACCACCACCACCACCACGGCGGCATGACAGTGGCGTATATT-3'

The 3' PCR primer included a XhoI restriction site and is shown below:

3'EcXhoHPPK SEQ ID NO: 33
5'-CGGCTCGAGTTACCATTTGTTTAATTTGT-3'

Using these primers, the 487 bp HPPK PCR product was amplified in a standard PCR amplification reaction, and an aliquot of the reaction was analyzed by agarose gel electrophoresis. A single band of approximately 519 bp that corresponded to the anticipated size was detected. The PCR amplification product was purified using a QIAquick PCR Purification Kit (QIAGEN Inc., Valencia, CA), and digested with the restriction enzymes NcoI and XhoI. The digested PCR product was ligated into NcoI- and XhoI-digested plasmid pBAR5, and an aliquot of the ligation reaction was used to transform TOP10 bacteria according to the manufacturer's protocols (Invitrogen Corp., Carlsbad, CA). Single colonies were isolated after plating on LBM agar plates containing carbenicillin. Several of the single colonies were grown overnight in 2 ml cultures of LBM containing carbenicillin, and plasmid DNA was isolated for DNA sequencing as previously described. The plasmid pBAR5/HisHPPK was shown to contain an open reading frame encoding the His(6)-HPPK gene having the following amino acid sequence set out in SEQ ID NO: 34.

3. HisHPPK Expression

Plasmid pBAR5/HisHPPK was used to transform the *E. coli* strain BL21(DE3)pLysS (Novagen Inc., Madison, WI) using standard methods. Transformants were selected after plating onto LBM plates containing both chloramphenicol and carbenicillin, to select for the plasmids pLysS and

pBAR5/HisHPPK respectively. Plasmid pLys is a plasmid that encodes T7 lysozome. The presence of lysozome aids cell lysis following a freeze-thaw cycle.

To initiate large-scale expression of HisHPPK, a 50 ml culture of BL21(DE3)pLysS containing pBAR5/HisHPPK was grown overnight at 30°C with shaking in LBM containing carbenicillin and chloramphenicol. The following day, 10 ml of the overnight culture was used to inoculate 2 liter flasks containing 500 ml of LBM supplemented with carbenicillin and chloramphenicol. The flasks were incubated at 37°C with shaking until the bacterial cultures reached an OD_{600} of approximately 0.6.

The plasmid pBAR5/HisHPPK contains an arabinose-inducible promoter upstream of the HisHPPK gene. Once the cultures reached appropriate density, arabinose was added to the cultures to a final concentration of 0.1% to induce HisHPPK expression, and the flasks were incubated at 37°C with shaking for another 2.5 hours. The bacteria were then harvested by centrifugation and the cell pellet from 1 liter of bacterial culture was resuspended in lysis buffer [50 mM Na_2HPO_4, pH 8, 50 mM imidazole, 10 mM β-mercaptoethanol, 0.5 M NaCl, and EDTA-free protease inhibitor cocktail tablets (Roche Molecular Biochemicals, Indianapolis, IN)] to a final volume of 35 ml. Each 35 ml bacterial suspension was transferred to a 50 ml polypropylene tube, snap frozen on dry ice, and then stored at -20°C.

4. Purification of HisHPPK

Each 35 ml aliquot was thawed on ice and lysed in a French press. To obtain a cleared lysate representing the soluble protein fraction, the lysate was centrifuged for 30 minutes at 20,000 x g, at 4°C. HisHPPK was purified using a two-step procedure.

First, bacterial proteins that bound the Ni-NTA agarose (QIAGEN Inc., Valencia, CA) nonspecifically were removed by incubating the cleared lysate with NTA agarose which had been previously treated with EDTA to remove associated Ni^{2+}cations. The 35 ml of cleared lysate was incubated batchwise with 1 ml of NTA agarose for approximately 1 hour at 4°C after which the NTA resin was

removed by centrifugation. HisHPPK was purified on Ni-NTA agarose according to the manufacturer's protocols (QIAGEN Inc., Valencia, CA). The isolated HisHPPK protein was resolved on a 12% Novex gel (Invitrogen Corp., Carlsbad, CA), the gel was fixed and stained with Coomassie brilliant blue under standard conditions, and the only protein identified in the HisHPPK preparation was a single species of about 19 kD in mass, which corresponds to the anticipated size of HisHPPK. The protein was dialyzed against 20 mM Tris, pH 8, aliquotted, and stored at -70°C.

5. Screening Assay for HPPK Activity

In order to identify small molecule inhibitors of HPPK, an assay for HPPK measuring the HPPK-dependent conversion of ATP to AMP as a by-product of the pyrophosphorylation of 6-hydroxymethyl-7,8-dihydropterin (HMDP) was employed [Shi *et al. J. Med. Chem.* 44:1364-1371 (2001)]. Elevated concentrations of both substrates (HMDP and ATP) were used in the assay to reduce the possibility of identifying substrate competitors. This reaction was modified for use in 96-well V-bottom polypropylene plates as follows

A master mix of the following composition was prepared containing 50 mM Hepes, pH 8.5, 100 µM HMDP (Schircks Laboratories, Jona, Switzerland), 10 mM $MgCl_2$, 35 µM adenosine triphosphate, and 10 ng of γ-labeled ^{32}P-ATP (Amersham Pharmacia Biotech, Arlington Heights, IL). An aliquot of the master mix was added to each well of the 96-well assay plate. Also added to the assay plate was 5 µl/well of the candidate inhibitor compound at a final screening concentration of 20 µM. Each candidate compound was diluted in DMSO prior to addition to the assay plate; and the final concentration of DMSO in the final assay mixture was 5%. The reaction was initiated by the addition of 100 ng of purified HisHPPK, and allowed to proceed for 15 minutes at 37°C. The reaction was stopped by the addition of an equal volume of 120 mM EDTA to each well. To resolve radiolabeled ATP from AMP, 2 µl of the reaction volume was spotted onto a PEI cellulose plate and the plate was developed with 0.3 M KH_2PO_4. The radioactivity of the plate was measured with a system Molecular Dynamics Storm 860 Phosphor imager system (Molecular Dynamics Storm, Sunnyvale, CA). The HPPK enzymatic activity in the

presence of compound was inferred from the percent conversion of radiolabeled ATP to AMP in duplicate test samples relative to a DMSO-only control reaction. Since nonspecific background in samples lacking substrate was less that 1%, and no correction was made. Approximately 2,520 compounds were screened, and approximately 58 compounds inhibited HPPK activity by 55% or greater, yielding a hit rate of 2.3%. These compounds were ranked for *in vitro* potency by IC50 determinations.

6. Effect of HPPK Antagonists on the Bacterial Growth of *E. coli* TolC

The minimal inhibitory concentration (MIC) required to inhibit the growth of *E. coli*, using a microtiter broth assay, was measured in order to determine the *in vivo* activity of the HPPK hits. The MIC is defined as the minimum concentration required to reduce growth 80% compared to DMSO-only controls. More specifically, the efficacy of these compounds was measured against an *E. coli* strain containing a mutation in the TolC gene. The TolC gene encodes a transperiplasmic efflux pump which facilitates the export of small molecules such as protein toxins and antibiotics from the bacterial cytosol [Andersen *et al.* Curr. Opin. Cell. Biol. 13:412-416 (2001).] Although, this mutation has no affect on the entry of compounds into the bacterium, some compounds prone to elimination via the efflux pumps may reach a higher intracellular concentration in the TolC mutant. All microtiter broth assays followed those protocols established by the National Committee for Clinical Laboratory Standards [*Methods for Dilution Antimicrobial Susceptibility Tests for Bacteria that Grow Aerobically*; approved standard-5th Edition. Vol. 20, No.2. NCCLS Guidelines. Wayne, Pennsylvania (2000).]

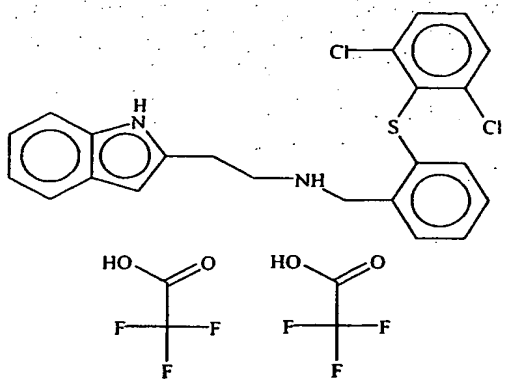
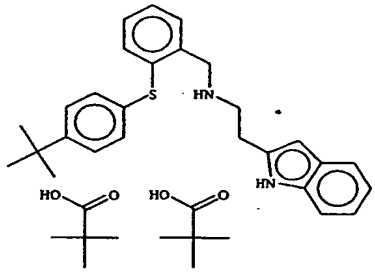
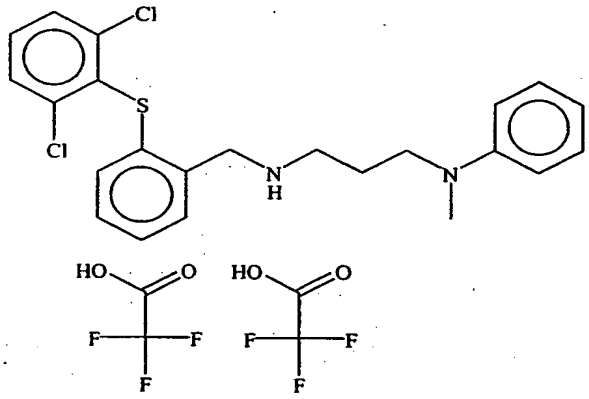
Microtiter broth assays were performed in Mueller-Hinton broth, which contains low thymidine levels. The presence of thymidine in bacterial media antagonizes the activity of the anti-folates trimethoprim and sulfamethoxazole, and likely antagonizes HPPK inhibitors as well.

Compounds were serially diluted two-fold in DMSO prior to addition to the microdilution plates. Each plate contained two controls: a serial dilution of trimethoprim provided a positive control for each plate, and a second row containing

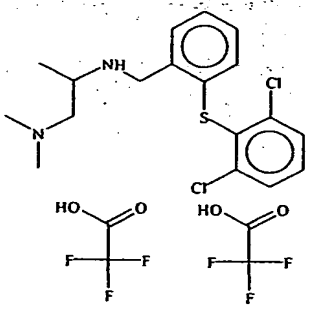
uninoculated Mueller-Hindon broth served as a sterility control for monitoring cross-contamination between wells. The inoculum density was approximately 10^5 bacteria/ml in a final volume of 100 µl. Plates were incubated for 16 hours before OD_{600} was measured.

The four compounds with the greatest activity in the MIC assays are shown in Table 9. The minimal inhibitory concentration of these compounds in E. coli TolC ranged from 0.1-12.5 µM. However, the MIC assays do not distinguish between bacteriostatic and bacteriocidal modes of action, nor do they determine if these compounds selectively inhibit HPPK *in vivo*. Experiments are underway to determine if these compounds have anti-folate activity and inhibit HPPK *in vivo*. It is well established that the activity of conventional anti-folates such trimethoprim and sulfamethoxazole are antagonized by the presence of thymidine in the bacterial medium [Amyes and Smith, J. Med. Microbiol. 7(2):143-153 (1973)]. Experiments to determine the MIC for each compound in Mueller-Hinton media alone, or following supplementation of the media with thymidine will be conducted. If the diarylsulfide compounds inhibit HPPK *in vivo*, then their activity should be attenuated in the presence of the folate end-product thymidine. Alternatively, these compounds can be analyzed for their ability to synergistically inhibit bacterial growth when paired with trimethoprim. Synergism would only occur if both the diarylsulfide compound and trimethoprim were acting on the same biochemical pathway. The combinatorial analysis of trimethoprim and a test compound are performed in a standard "checkerboard" study where these compounds are cross-titrated and analyzed for their effect on bacterial growth in a microtiter broth assay as previously described [Eliopoulos and Moellering, Jr., *Antimicrobial Combinations*, pp. 330-393, *in* Antibiotics in Laboratory Medicine, 4th Edition.(V. Lorian ed., 1996)].

TABLE 9

AAD 
AAE 
AAF 

AAG



Example 20
Assays for the Identification of ftsZ Inhibitors

FtsZ is the product of an essential bacterial gene that is involved in cell division. FtsZ binds and hydrolyzes GTP, and when bound to GTP it forms long, linear polymers. The GTP-dependent polymerization of ftsZ is related to its function in bacterial cell division. During septation, ftsZ forms a ring to define the plane of cell division. Cells lacking ftsZ can not undergo septation, do not divide and die. FtsZ is highly conserved (approximately 60%) throughout the bacterial kingdom. Accordingly, ftsZ inhibitors could represent broad-spectrum antibiotics with a novel mechanism of action. The atomic structure of ftsZ, as determined by x-ray diffraction, shows that it is an alpha/beta protein [Nogales *et al.*, (1998) Nature Structural Biology 5:451-458]. The most similar structural relative to ftsZ is the eukaryotic protein tubulin, which is a GTP-binding and hydrolyzing protein that also polymerizes to form microtubules with an essential role in the segregation of organelles and chromosomes during cell division.

A polymerization assay for the identification of ftsZ inhibitors that can be performed in microtiter wells has been devised. The polymerization assay is an adaptation of a tubulin polymerization assay [Bollag *et al.*, Cancer Research 55:2325-2333 (1995)], and involves the reversible polymerization of ftsZ in a GTP-dependent fashion.

In the presence of GTP and 10mM $CaCl_2$, 5 nm ftsZ linear polymers assemble into higher order polymers [Yu *et al.*, EMBO 16:5455-5463 (1997)] that are large enough to be trapped by a 0.2μm filter. The protein that is retained on the filter can be stained and detected in a colorimetric assay. A reaction consisting of 300 μg/ml of ftsZ polymerized by 100μM GTP was screened against candidate ftsZ inhibitors at 10μM.

An alternative assay that may be more sensitive was also devised. In this assay, 100μg/ml ftsZ was incubated with 0.5 μM ^{32}P-γ-GTP. The GTPase activity of ftsZ liberates $^{32}PO_4$. By terminating the reaction with 25 mg/ml activated charcoal in 100mM NaH_2PO_4 and centrifuging the product, the remaining ^{32}P-γ-GTP is trapped by the charcoal. Accordingly, the $^{32}PO_4$ that remains in the supernatant can be measured, providing a measurement of GTPase inhibition. This screening

assay may better identify ftsZ inhibitors because it is significantly more sensitive to inhibition by GDP than the polymerization screen described above (IC50 of 8 μM vs. 250 μM).

Example 21
Screening Assay for ENR Inhibitors

An assay to screen for ENR inhibitors, using non-radioactive high purity NADH, was developed. The isolation of ENR is described by Baldock *et al.*, Science 274:2107 (1996). Briefly, ENR catalyzes the conversion of NADH and crotonyl-CoA to form NAD+ and fatty acyl-CoA, and the assay measures the amount of NAD+ produced in a second reaction wherein luciferase converts NAD+ to NADH. Light emission from the luciferase reaction is proportional to the amount of NAD+ produced in the initial reaction. A candidate inhibitor compound is added to the ENR reaction, and if the candidate inhibits ENR activity, the amount of light detected in the luciferase reaction is decreased.

The assay was carried out as follows. Twenty μl of 30 μM NADH (Boehringer Manneheim) in 20mM Hepes containing 6 ng/μl ENR or a total of 120 ng per well and 20 μl of 10 μM candidate inhibitor compound in DMSO were added to a 96 well flat bottom optical plate. Twenty μl of 300 μM crotonyl-CoA (Sigma, C6146) was subsequently added to initiate the reaction. Triclosan was used as a control inhibitor and was included on each plate to verify inhibition. In the screening assay, triclosan inhibits with an IC50 of about 1 μM.

The reaction was allowed to continue for approximately ten minutes, corresponding to about 30 percent of the way to completion. Accordingly, the concentration of NAD+ should be approximately 3 μM after ten minutes.

Thirty μl of 160mM HCl was added to the system to bring the pH of the reaction mixture below 2 and remove remaining NADH substrate. The reaction mixture was incubated for one minute following acid addition so that the remaining NADH decomposes to ADP-ribose and nicotinamide. NAD+ is substantially unaffected by the addition of strong acid.

After the one minute period referenced immediately above, 110 μl of a NADH regeneration/luciferase solution comprising alcohol dehydrogenase,

ethanol. FMN, FMN oxidoreductase, decanal and bacterial luciferase was added to the reaction mixture.

More specifically, the NADH regeneration/luciferase solution was prepared in 110 µl of a buffer solution containing 300mM Tris (using a stock 1 M, pH 7.5 solution), 0.26 % by weight bis(trimethylsilyl)acetamide ("BMA"), 0.65 mM EDTA, and 18 mM KCl. To this solution, 0.67 µl of decanal (Sigma, D7384, 98% purity) was added for every 10 ml of solution to yield a final solution having decanal concentration of approximately 200 µM. Sufficient FMN (Sigma F8399) was added to provide a final solution having a FMN concentration of approximately 2 µM. Similarly, sufficient ethanol (200 proof) was added such that the final solution has an ethanol concentration of approximately 100 µM. After adding all of these reagents to the solution, it was vortexed vigorously.

To this solution, 1.08 µl of NADH:FMN oxidoreductase (Roche, 476 480) was added for each 10 ml of solution, to yield a final solution having a concentration of 1.25 units per liter. Bacterial luciferase (Roche, 476 498) was added to yield a solution having approximately 4.5 µg/ml. Similarly, alcohol dehydrogenase was added to provide a solution having a final concentration of 0.7 units per ml. After adding these reagents, the mixture was mixed gently by inversion.

Approximately 100 compounds that inhibit ENR activity were identified in this assay. About 50 of these compounds exhibited significant inhibitory activity in a radiomimetric ENR assay. In this assay, twenty µl of 30 µM ^{32}P-NADH in 20mM Hepes was incubated with 120 ng of ENR per well and 20 µl of 10 µM candidate inhibitor compound in DMSO. Twenty µl of 300 µM crotonyl-CoA (Sigma, C6146) was subsequently added to initiate the reaction. The products of the reaction include 32P-NAD. The reactant 32P-NADH and the product 32P-NAD are separated from each other by thin layer chromatography on PEI-cellulose in 1M LiCl, and visualized by autoradiography. The extent of the reaction is determined by the conversion of NADH to NAD. These compounds were further tested for inhibition of *E. coli* growth.

A permeable bacterial strain (AB734 TN10::tolC) was used in the screening method to maximize the ability of the compounds to cross the gram negative cell wall. Assays were conducted in accordance with the NCCLS protocols referenced herein.

It was of further interest to determine whether those compounds with antimicrobial activity worked in a ENR-dependent fashion. Two strains of the permeable tolC strain were constructed. In the first strain, the ENR protein was overexpressed by placing it under control of its own promoter on a moderate copy number plasmid, and the second strain served as a control including only the plasmid. A candidate compound that targets ENR should be much less active against the first strain described above because the target is substantially overexpressed. For example, the MIC for triclosan shifts from 31 to 1000 ng/ml when tested against the first strain. Similarly, compound 325084 had a shift in MIC from 25 μM to greater than 100 μM, suggesting that this compound exerts its antimicrobial action by virtue of inhibiting ENR during bacterial growth. The results do not distinguish between the possibilities that compound 325084 and triclosan act on ENR at the same or distinct sites on the enzyme. However, because no other compound showed a similar shift in MIC, it is believed that these other compounds probably inhibit bacterial growth through a different mechanism. Nonetheless, compounds 325085 and 325086 have structures similar to compound 325084, and also demonstrated some activity against ENR.

In order to determine if compound 325084 and triclosan act at the same or different ENR sites, recombinant ENR is produced which (i) includes a glycine to valine substitution at residue 93, (ii) retains enzymatic activity, and (iii) is insensitive to triclosan. Compounds that are identified as ENR inhibitors are then assayed using both wild type and mutant ENR and compounds that show little or no inhibitory activity against the mutant ENR form are probably acting at the active site of ENR and may be discarded. Alternatively, compounds which inhibit the mutant enzyme and the wild type form may be acting at an allosteric site and will be studied further.

While the present invention has been described in terms of specific embodiments, it is understood that variations and modifications will occur to those skilled in the art. For example, with respect to the compounds disclosed herein, it should be understood that the substitution of one halogen substituent for another, different halogen substituent is within the scope of the present invention. Accordingly, only such limitations as appear in the appended claims should be placed on the present invention.

What is claimed is:

1. A method of modulating binding interaction between a first molecule which is not LFA-1 or an I domain-containing fragment thereof, and a binding partner molecule, said first molecule comprising an α/β domain structure, said α/β structure comprising an allosteric regulatory site, said method comprising the step of contacting said first molecule with an allosteric effector molecule that interacts with said allosteric regulatory site and promotes a conformation in a ligand binding domain of said α/β structure that modulates binding between said first molecule and said binding partner molecule.

2. A method of modulating binding interaction between a first molecule which is not LFA-1 or an I domain-containing fragment thereof, and a binding partner molecule, said first molecule comprising an α/β domain structure, said α/β structure comprising an allosteric regulatory site, said method comprising the step of contacting said first molecule with an allosteric effector molecule, said allosteric effector molecule comprising a diaryl compound, said diaryl compound interacting with said allosteric regulatory site and promoting a conformation in a ligand binding domain of said α/β structure that modulates binding between said first molecule and said binding partner molecule.

3. A method of modulating binding interaction between a first molecule which is not LFA-1 or an I domain-containing fragment thereof, and a binding partner molecule, said first molecule comprising an α/β domain structure, said α/β structure comprising an allosteric regulatory site, said method comprising the step of contacting said first molecule with an allosteric effector molecule, said allosteric effector molecule selected from the group consisting of diaryl sulfide compounds and diarylamide compounds, said allosteric effector molecule interacting with said allosteric regulatory site and promoting a conformation in a ligand binding domain of said α/β structure that modulates binding between said first molecule and said binding partner molecule.

4. The method of claim 1, 2, or 3 wherein said first molecule comprises a Rossmann fold structure, said Rossmann fold structure comprising said allosteric regulatory site.

5. The method of claim 4 wherein said Rossmann fold structure in said first molecule comprises a β sheet having β sheet strands positioned in a 321456 or 231456 orientation.

6. The method of claim 4 wherein said Rossmann fold structure in said first molecule comprises a β sheet having β sheet strands positioned in a 3214567 orientation.

7. The method of claim 4 wherein said Rossmann fold structure in said first molecule comprises a β sheet having β sheet strands positioned in a 32145 orientation.

8. The method of claim 1, 2, or 3 wherein said first molecule comprises an I domain structure.

9. The method of claim 1, 2, or 3 wherein said first molecule comprises an A domain structure.

10. A method of modulating binding interaction between a first molecule and a binding partner molecule, said first molecule having an amino acid sequence which exhibits less than about 90% identity to the LFA-1 I domain amino acid sequence set out in FIGURE 1, said first molecule comprising an α/β structure, said α/β domain structure comprising an allosteric regulatory site, said method comprising the step of contacting said first molecule with an allosteric effector molecule that interacts with said allosteric regulatory site and promotes a conformation in a ligand binding domain of said α/β structure that modulates binding between said first molecule and said binding partner molecule.

11. A method of modulating binding interaction between a first molecule and a binding partner molecule, said first molecule having an amino acid sequence which exhibits less than about 90% identity to the LFA-1 I domain amino acid sequence set out in FIGURE 1, said first molecule comprising an α/β structure, said α/β domain structure comprising an allosteric regulatory site, said method comprising the step of contacting said first molecule with an allosteric effector molecule, said allosteric effector molecule comprising a diaryl compound, said diaryl compound interacting with said allosteric regulatory site and promoting a conformation in a ligand binding domain of said α/β structure that modulates binding between said first molecule and said binding partner molecule.

12. A method of modulating binding interaction between a first molecule and a binding partner molecule, said first molecule having an amino acid sequence which exhibits less than about 90% identity to the LFA-1 I domain amino acid sequence set out in FIGURE 1, said first molecule comprising an α/β domain structure, said α/β structure comprising an allosteric regulatory site, said method comprising the step of contacting said first molecule with an allosteric effector molecule, said allosteric effector molecule selected from the group consisting of diaryl sulfide compounds and diarylamide compounds, said allosteric effector molecule interacting with said allosteric regulatory site and promoting a conformation in a ligand binding domain of said α/β structure that modulates binding between said first molecule and said binding partner molecule.

13. The method of claim 10, 11, or 12 wherein said first molecule has an amino acid sequence that exhibits a percent identity with respect to the LFA-1 I domain amino acid sequence less than about 40%, about 45%, about 50%, about 55%, about 60%, about 65%, about 70%, about 75%, about 80%, about 85%, or about 90%.

14. The method of claim 10, 11, or 12 wherein said first molecule comprises a Rossmann fold structure, said Rossmann fold structure comprising an allosteric regulatory site.

15. The method of claim 14 wherein said first molecule has an amino acid sequence that exhibits a percent identity with respect to the LFA-1 I domain amino acid sequence less than about 40%, about 45%, about 50%, about 55%, about 60%, about 65%, about 70%, about 75%, about 80%, about 85%, or about 90%.

16. The method of claim 14 wherein said Rossmann fold structure in said first molecule comprises a β sheet having β sheet strands positioned in a 321456 or 231456 orientation.

17. The method of claim 16 wherein said first molecule has an amino acid sequence that exhibits a percent identity with respect to the LFA-1 I domain amino acid sequence less than about 40%, about 45%, about 50%, about 55%, about 60%, about 65%, about 70%, about 75%, about 80%, about 85%, or about 90%.

18. The method of claim 14 wherein said Rossmann fold structure in said first molecule comprises a β sheet having β sheet strands positioned in a 3214567 orientation.

19. The method of claim 18 wherein said first molecule has an amino acid sequence that exhibits a percent identity with respect to the LFA-1 I domain amino acid sequence less than about 40%, about 45%, about 50%, about 55%, about 60%, about 65%, about 70%, about 75%, about 80%, about 85%, or about 90%.

20. The method of claim 14 wherein said Rossmann fold structure in said first molecule comprises a β sheet having β sheets strands positioned in a 32145 orientation.

21. The method of claim 20 wherein said first molecule has an amino acid sequence that exhibits a percent identity with respect to the LFA-1 I domain amino acid sequence less than about 40%, about 45%, about 50%, about 55%, about 60%, about 65%, about 70%, about 75%, about 80%, 85%, or about 90%.

22. The method of claim 10, 11, or 12 wherein said first molecule comprises an I domain structure.

23. The method of claim 23 wherein said first molecule has an amino acid sequence that exhibits a percent identity with respect to the LFA-1 I domain amino acid sequence less than about 40%, about 45%, about 50%, about 55%, about 60%, about 65%, about 70%, about 75%, about 80%, about 85%,or about 90%.

24. The method of claim 11, 11, or 12 wherein said first molecule comprises an A domain structure.

25. The method of claim 24 wherein said first molecule has an amino acid sequence that exhibits a percent identity with respect to the LFA-1 I domain amino acid sequence less than about 40%, about 45%, about 50%, about 55%, about 60%, about 65%, about 70%, about 75%, about 80%, about 85%, or about 90%.

26. The method of any one of claims 1-3, 5-7, 10-12, 15-21, 23 or 25 wherein the modulator promotes a conformation in the ligand binding domain of said

first molecule that increases binding between said first molecule and said binding partner molecule.

27. The method of claim 26 wherein the increase in binding between the first molecule and the second molecule results in increased enzymatic activity of the first molecule.

28. The method of any one of claims 1-3, 5-7, 10-12, 15-21, 23 or 25 wherein the modulator promotes a conformation in the ligand binding domain of said first molecule that decreases binding between said first molecule and said binding partner molecule.

29. The method of claim 28 wherein the decrease in binding between the first molecule and the second molecule results in decreased enzymatic activity of the first molecule.

30. The method of any one of claims 1-3, 5-7, 10-12, 15-21, 23 or 25 wherein the first molecule is selected from the group consisting of the proteins set forth in Table 1.

31. The method of claim 30 wherein the first molecule is a eukaryotic molecule.

32. The method of claim 30 wherein the first molecule is a human molecule.

33. The method of claim 30 wherein the first molecule is a prokaryotic molecule.

34. The method of claim 30 wherein the first molecule is a bacterial molecule.

35. The method of claim 30 wherein the first molecule is selected from the group consisting of $\alpha_M\beta_2$, complement protein C2, complement protein Factor B, $\alpha_E\beta_7$, $\alpha_4\beta_7$, $\alpha_V\beta_3$, $\alpha_4\beta_1$, $\alpha_d\beta_2$,von Willebrand factor, Rac-1, HPPK, ftsZ, and ENR.

36. The method of claim 35 wherein the first molecule is $\alpha_M\beta_2$ and the binding partner protein is fibrinogen.

37. The method of claim 35 wherein the first molecule is $\alpha_M\beta_2$ and the binding partner protein is iC3b.

38. The method of claim 35 wherein the first molecule is $\alpha_E\beta_7$ and the binding partner protein is E-cadherin.

39. The method of claim 35 wherein the first molecule is $\alpha_4\beta_7$ and the binding partner protein is MAdCAM-1.

40. The method of claim 35 wherein the first molecule is $\alpha_V\beta_3$ and the binding partner protein is vitronectin.

41. The method of claim 35 wherein the first molecule is $\alpha_4\beta_1$ and the binding partner protein is VCAM.

42. The method of claim 35 wherein the first molecule is $\alpha_d\beta_2$ and the binding partner protein is VCAM.

43. The method of claim 35 wherein the first molecule is von Willebrand factor and the binding partner protein is gpIb.

44. The method of claim 35 wherein the first molecule is complement protein C2 and the binding partner protein is complement protein C4b.

45. The method of claim 35 wherein the first molecule is complement protein Factor B and the binding partner protein is complement protein C3b.

46. The method of claim 35 wherein the first molecule is Rac-1 and the binding partner is GTP.

47. The method of claim 35 wherein the first molecule is HPPK and the binding partner is ATP or HMDP.

48. The method of claim 35 wherein the first molecule is ftsZ and the binding partner is GTP.

49. The method of claim 35 wherein the first molecule is ENR and the binding partner is NADH.

ICOS Corporation
MATERIALS AND METHODS TO MODULATE LIGAND BINDING/ENZYMATIC ACTIVITY OF ALPHA BETA PROTEINS CONTAINING AN ALLOSTERIC REGULATORY SITE
Attorney Docket No. 27866/36470A

Figure 1 - LFA-1 I Domain Amino Acid Sequence

GNVDLVFLFD GSMSLQPDEF QKILDFMKDV MKKLSNTSYQ FAAVQFSTSY KTEFDFSDYV
KWKDPDALLK HVKHMLLLTN TFGAINYVAT EVFREELGAR PDATKVLIII TDGEATDSGN
IDAAKDIIRY IIGIGKHFQT KESQETLHKF ASKPASEFVK ILDTFEKLKD LFTELQKKIY

FIGURE 2 - LFA-1 Amino Acid Sequcene

MKDSCITVMA MALLSGFFFF APASSYNLDV RGARSFSPPR AGRHFGYRVL QVGNGVIVGA
PGEGNSTGSL YQCQSGTGHC LPVTLRGSNY TSKYLGMTLA TDPTDGSILA CDPGLSRTCD
QNTYLSGLCY LFRQNLQGPM LQGRPGFQEC IKGNVDLVFL FDGSMSLQPD EFQKILDFMK
DVMKKLSNTS YQFAAVQFST SYKTEFDFSD YVKWKDPDAL LKHVKHMLLL TNTFGAINYV
ATEVFREELG ARPDATKVLI IITDGEATDS GNIDAAKDII RYIIGIGKHF QTKESQETLH
KFASKPASEF VKILDTFEKL KDLFTELQKK IYVIEGTSKQ DLTSFNMELS SSGISADLSR
GHAVVGAVGA KDWAGGFLDL KADLQDDTFI GNEPLTPEVR AGYLGYTVTW LPSRQKTSLL
ASGAPRYQHM GRVLLFQEPQ GGGHWSQVQT IHGTQIGSYF GGELCGVDVD QDGETELLLI
GAPLFYGEQR GGRVFIYQRR QLGFEEVSEL QGDPGYPLGR FGEAITALTD INGDGLVDVA
VGAPLEEQGA VYIFNGRHGG LSPQPSQRIE GTQVLSGIQW FGRSIHGVKD LEGDGLADVA
VGAESQMIVL SSRPVVDMVT LMSFSPAEIP VHEVECSYST SNKMKEGVNI TICFQIKSLY
PQFQGRLVAN LTYTLQLDGH RTRRRGLFPG GRHELRRNIA VTTSMSCTDF SFHFPVCVQD
LISPINVSLN FSLWEEEGTP RDQRAQGKDI PPILRPSLHS ETWEIPFEKN CGEDKKCEAN
LRVSFSPARS RALRLTAFAS LSVELSLSNL EEDAYWVQLD LHFPPGLSFR KVEMLKPHSQ
IPVSCEELPE ESRLLSRALS CNVSSPIFKA GHSVALQMMF NTLVNSSWGD SVELHANVTC
NNEDSDLLED NSATTIIPIL YPINILIQDQ EDSTLYVSFT PKGPKIHQVK HMYQVRIQPS
IHDHNIPTLE AVVGVPQPPS EGPITHQWSV QMEPPVPCHY EDLERLPDAA EPCLPGALFR
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SEQUENCE LISTING

<110> ICOS CORPORATION

<120> MATERIALS AND METHODS TO MODULATE LIGAND BINDING/ENZYMATIC ACTIVITY OF ALPHA/BETA PROTEINS CONTAINING AN ALLOSTERIC REGULATORY SITE

<130> 27866/36470A

<140> To be determined
<141> Filed herewith

<150> US 60/239,750
<151> 2000-10-12

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Leu Asn Ala Ala Val Ala Leu Glu Thr Ser Leu Ala Pro Glu Glu Leu
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60/239,750 12 October 2000 (12.10.2000) US

(71) Applicant: ICOS CORPORATION [US/US]; 22021 20th Avenue, S.E., Bothell, WA 98021 (US).

(72) Inventor: STAUTON, Donald, E.; 6502 113th Avenue NE, Kirkland, WA 98033 (US).

(74) Agent: WILLIAMS, Joseph, A., Jr.; Marshall, Gerstein & Borun, 6300 Sears Tower, 233 South Wacker Drive, Chicago, IL 60606 (US).

(81) Designated States *(national)*: AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, PH, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, UZ, VN, YU, ZA, ZW.

(84) Designated States *(regional)*: ARIPO patent (GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, TR), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NE, SN, TD, TG).

Published:
— *with international search report*

(88) Date of publication of the international search report:
13 March 2003

For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

WO 02/031511 A3

(54) Title: MODULATION OF LIGAND BINDING/ENZYMATIC ACTIVITY OF ALPHA BETA PROTEINS

(57) Abstract: Methods of modulating binding between an α/β protein and a binding partner are provided, along with methods of identifying modulators and their use.

INTERNATIONAL SEARCH REPORT

Int[redacted]nal Application No
PCT/US 01/32047

A. CLASSIFICATION OF SUBJECT MATTER
IPC 7 G01N33/68

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
IPC 7 G01N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)
EPO-Internal, PAJ, WPI Data, BIOSIS, MEDLINE

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category °	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	LEE J-O ET AL: "CRYSTAL STRUCTURE OF THE A DOMAIN FROM THE ALPHA SUBUNIT OF INTEGRIN CR3 (CD11B/CD18)" CELL, CELL PRESS, CAMBRIDGE, NA, US, vol. 80, 24 February 1995 (1995-02-24), pages 631-638, XP002920554 ISSN: 0092-8674 the whole document --- -/--	1-49

[X] Further documents are listed in the continuation of box C.

[ ] Patent family members are listed in annex.

° Special categories of cited documents:

"A" document defining the general state of the art which is not considered to be of particular relevance

"E" earlier document but published on or after the international filing date

"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

"&" document member of the same patent family

Date of the actual completion of the international search	Date of mailing of the international search report
29 October 2002	06/11/2002

Name and mailing address of the ISA	Authorized officer
European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Tx. 31 651 epo nl, Fax: (+31-70) 340-3016	Moreno de Vega, C

Form PCT/ISA/210 (second sheet) (July 1992)

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	JONES J I ET AL: "LIGAND OCCUPANCY OF THE ALPHAV BETA3 INTEGRIN IS NECESSARY FOR SMOOTH MUSCLE CELLS TO MIGRATE IN RESPONSE TO INSULIN-LIKE GROWTH FACTOR I" PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES OF USA, NATIONAL ACADEMY OF SCIENCE. WASHINGTON, US, vol. 93, 1 March 1996 (1996-03-01), pages 2482-2487, XP002058895 ISSN: 0027-8424	1,8,10, 22,26, 28, 30-32, 35,40
Y	the whole document	1-49

Y	OXVIG CLAUS LU C ET AL: "Conformational changes in tertiary structure near the ligand binding site of an integrin I domain" PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES OF USA, NATIONAL ACADEMY OF SCIENCE. WASHINGTON, US, vol. 96, no. 5, 2 March 1999 (1999-03-02), pages 2215-2220, XP002209413 ISSN: 0027-8424 the whole document	1-49

Y	NOLTE M ET AL: "Crystal structure of the alpha1beta1 integrin I-domain: insights into integrin I-domain function" FEBS LETTERS, ELSEVIER SCIENCE PUBLISHERS, AMSTERDAM, NL, vol. 452, no. 3, 11 June 1999 (1999-06-11), pages 379-385, XP004259786 ISSN: 0014-5793 the whole document	1-49

Y	HUTH J R ET AL: "NMR AND MUTAGENESIS EVIDENCE FOR AND I DOMAIN ALLOSTERIC SITE THAT REGULATES LYMPHOCYTE FUNCTION-ASSOCIATED ANTIGEN 1 LIGAND BINDING" PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES OF USA, NATIONAL ACADEMY OF SCIENCE. WASHINGTON, US, vol. 97, no. 10, 9 May 2000 (2000-05-09), pages 5231-5236, XP000938664 ISSN: 0027-8424 cited in the application the whole document	1-49

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INTERNATIONAL SEARCH REPORT

Ir [redacted] ional Application No
PCT/US 01/32047

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category °	Citation of document, with indication,where appropriate, of the relevant passages	Relevant to claim No.
Y,P	LU CHAFEN ET AL: "An isolated, surface-expressed I domain of the integrin alphaLbeta2 is sufficient for strong adhesive function when locked in the open conformation with a disulfide bond" PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES OF USA, NATIONAL ACADEMY OF SCIENCE. WASHINGTON, US, vol. 98, no. 5, 27 February 2001 (2001-02-27), pages 2387-2392, XP002209618 ISSN: 0027-8424 the whole document	1-49
Y,P	COUSIN, J.: "Integrin crystal structure solved" SCIENCE, no. 293, 7 September 2001 (2001-09-07), pages 1743-46, XP001106538 cited in the application the whole document	1-49

Form PCT/ISA/210 (continuation of second sheet) (July 1992)

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US 01/32047

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☐ Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:

2. ☒ Claims Nos.: –
because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:
see FURTHER INFORMATION sheet PCT/ISA/210

3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

1. ☐ As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.

2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.

3. ☐ As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:

4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

☐ The additional search fees were accompanied by the applicant's protest.

☐ No protest accompanied the payment of additional search fees.

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

Continuation of Box I.2

Present claims 1-34 relate to an extremely large number of possible methods. Support within the meaning of Article 6 PCT and disclosure within the meaning of Article 5 PCT is to be found, however, for only a very small proportion of the methods claimed. In the present case, the claims so lack support, and the application so lacks disclosure, that a meaningful search over the whole of the claimed scope is impossible. Consequently, the search has been carried out for those parts of the claims which appear to be supported and disclosed, namely those parts relating to the methods using the ligands disclosed in claims 35-49.

The applicant's attention is drawn to the fact that claims, or parts of claims, relating to inventions in respect of which no international search report has been established need not be the subject of an international preliminary examination (Rule 66.1(e) PCT). The applicant is advised that the EPO policy when acting as an International Preliminary Examining Authority is normally not to carry out a preliminary examination on matter which has not been searched. This is the case irrespective of whether or not the claims are amended following receipt of the search report or during any Chapter II procedure.